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INTERNATIONAL JOURNAL OF ADVANCED RESEARCH

RESEARCH ARTICLE

Acute toxicity prediction of synthetic and natural preservatives in rat by using QSAR modeling software

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Manuscript Info

Abstract

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Manuscript History:

Received: 15 May 2015 Final Accepted: 22 June 2015 Published Online: July 2015

Key words:

QSAR, preservatives, T.E.S.T. software, acute toxicity, rat

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Soumendra Nath Talapatra Preservatives are chemicals that prevent spoiling the foods from bacteria and fungi. These chemicals are of artificial or natural origin. The present study aims acute toxicity prediction as LD₅₀ (median lethal dose) values of synthetic/artificial and natural preservatives in rat by oral exposure. The predictions of LD_{50} values were determined by using T.E.S.T. (Toxicity Estimation Software Tool) for these common preservatives used in raw and/or processed foods. Among the selected 16 synthetic and natural preservatives, predictive LD₅₀ values by T.E.S.T. were obtained only in 8 artificial compounds while for natural preservatives, out of 16 compounds 15 nos. were obtained. The predicted LD₅₀ values for synthetic compounds viz. benzoic acid, ethyl para-hydroxybenzoate, lactic acid, propionic acid, sorbic acid, butylated hydroxytoluene, butylated hydroxyanisole and formaldehyde were observed but potassium sorbate, sodium benzoate, calcium benzoate, sodium propionate, disodium EDTA, sodium gallate, nitrite and sulphite unable to predict the LD₅₀ values due to unidentified CAS no. in database while for natural compounds viz. caffeic acid, cinnamic acid, ferulic acid, gallic acid, oleuropein, thymol, eugenol, ascorbic acid, tartaric acid, malic acid, fumaric acid, tocopherol acetate, carvacrol, citral and allin were obtained. It was observed from the prediction by the T.E.S.T. that synthetic preservatives are moderately toxic except formaldehyde and propionic acid, these 2 are very toxic while natural preservatives are only moderately toxic and lactic acid is slightly toxic. This software helps to predict of LD₅₀ values by easy screening. It is also suggested to compare the predicted data with other available related softwares and natural preservatives may suitable compared to artificial preservatives.

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INTRODUCTION

Preservatives are chemical compounds that can originate from both artificial and natural source. The functions of artificial and natural preservatives in food substances are mainly killing the growth of microorganisms such as bacteria and fungi (Soeda et al., 1966; CAL, 1988; Cutler, 1995; Puupponen et al., 2001; Erasto et al., 2004; Souza et al., 2005; Mohanka and Priyanka, 2014). The artificial or synthetic preservatives have formed by several chemical reactions but the natural preservatives can directly be obtained from plant and/or animal origin, which are known as bioactive compounds. It has already been established the physical and chemical nature, mode of actions etc. of both artificial as well as natural chemical compounds used as preservative in food to prevent spoilage from bacteria and

fungi (Close and Nielsen, 1976; Chapman, 1998; Nararasimhan et al., 2003; Narasimhan et al., 2004; Ou and Kwok, 2004; Bhullar et al., 2013; Mohanka and Priyanka, 2014).

In new research era, quantitative structure–activity relationship (QSAR) is logic based mathematical simulation of the structure-derived features of a chemical compound and their relation with biological or physicochemical activity already established for one or more than one chemical as preservative (Narasimhan and Dhake, 2006; Narasimhan et al., 2006; Chaudhary et al., 2008; Narasimhan et al., 2009; Roy et al., 2011; Sarova et al., 2011; Mahiwal et al., 2012). QSAR modeling helps to predict several biological activity viz. acute toxicity, mutagenicity, developmental toxicity etc. or inhibitory versus non-inhibitory activities of compound(s) prior to bioassay. The established molecular descriptors have been used to determine the exact prediction of chemical(s). Generally the molecular descriptors are used on the basis of three parameters viz. thermodynamic, steric and electronic (Choplin, 2005; Valentina et al., 2009) in QSAR modeling. In T.E.S.T., the parameters also include E-state values and E-state counts, constitutional descriptors, topological descriptors, walk and path counts, connectivity, information content, 2D autocorrelation, Burden eigenvalue, molecular property (such as the octanol-water partition coefficient), Kappa, hydrogen bond acceptor/donor counts, molecular distance edge, and molecular fragment counts (Cassano et al., 2010; USEPA, 2012).

The bioassay with animals for any compound is difficult to get data within a short duration, need more expenditure and time-consuming, presently some restrictions to use animal species by the regulatory authorities. In this context, the toxicity prediction has been postulated through QSAR along with statistical modeling (Zhu et al., 2009), which is a computer based suitable technique (Worth et al., 2007; Patil et al., 2014). Scattered works on toxicity prediction by using QSAR modeling have been reported for preservatives from artificial as well as natural origin (Narasimhan et al., 2003; 2004; 2006; Chaudhary et al., 2008; Venkataramana et al., 2011; Mahiwal et al., 2012).

It has already been recommended several toxicity prediction softwares by many researchers (Pallas, Compu Drug International Inc., 2000; TOPKAT, 2004; Talete, 2006; User manual (ADMET), Simulation Plus Inc, 2011; USEPA, 2012). Among all these softwares, the T.E.S.T. (Toxicity Estimation Software Tool) by USEPA (2012) is a non-commercial software and easy to operate for obtaining predictive values within short period of time (Ruiz et al., 2002; Talapatra et al., 2015). In this package, it was found QSAR modeling to calculate the toxicity of chemical(s) using suitable molecular descriptors along with statistical interpretation.

In the T.E.S.T software, multiple prediction methodologies have been incorporated, which has higher confidence level in the toxicity prediction (the values for predicted toxicities are closely related from different methods). In addition, particular QSAR approaches depend upon the confidence at higher level by individual experience of researchers (USEPA, 2012).

In this present study an attempt has been made to predict acute toxicity as LD_{50} values of artificial and natural chemicals found in preservatives in the rat orally exposed through QSAR modeling software package T.E.S.T.

MATERIALS AND METHODS

In the present study, established 16 types of synthetic and natural preservatives were selected based on the usage in raw and processed food items. The present study was emphasized to estimate the LD_{50} values of synthetic and natural preservatives through QSAR modeling software package (T.E.S.T., Verson 4.1) in rat oral exposure (US EPA, 2012) and this easy screening software was used for acute toxicity prediction. The value for acute toxicity study was tabulated after obtaining predictive value of individual compound from T.E.S.T. software. It was studied the LD_{50} data from a consensus method, which is basically the average predicted LD_{50} value calculated from other inbuilt QSAR methodologies viz. hierarchical clustering method, the FDA MDL method and nearest neighbor methods (Martin et al., 2008; USEPA, 2012). In T.E.S.T., the structure of studied chemical can only be visualized when entering CAS no. of individual chemical by clicking on calculate option. The predicted value of consensus method can only be obtained after processing the software. This software is programmed by the inbuilt of 7,420 chemicals' database (Martin et al., 2008) and is only based on 2-dimensional molecular descriptors of 797 nos. (Zhu et al., 2009; USEPA, 2012).

In T.E.S.T., it is prescribed that the R^2 (correlation coefficient) value if shows greater than 60% then the prediction data is reliable and the predictive ability of each of the QSAR methodologies is evaluating by statistical external validation (Golbraikh and Tropsha, 2002; Golbraikh et al., 2003; Gramatica and Pilutti, 2004; Zhu et al., 2009). According to Zhu et al. (2009), the applicability domain (AD) is calculated for each method to justify exact prediction. The AD is based on the correct validation of predictive activity in T.E.S.T. (USEPA, 2012). It was known that the QSAR modeling can only predict the potential toxicity of any chemical but the confidence level in such predictions can vary (USEPA, 2012). However, in the QSAR model ADs, the features and limitations can be

understood in detail after suitable interpretation of predictions (Golbraikh and Tropsha, 2002; Golbraikh et al., 2003; Netzeva et al., 2005; Schultz et al., 2007; Tropsha and Golbraikh, 2007; Tetko et al., 2008; Zhu et al., 2009; Roy et al., 2011; Ruiz et al., 2012). The predictive value for acute toxicity with special reference to LD_{50} values in rat after orally exposed to artificial and natural preservatives were only determined by using QSAR modeling T.E.S.T. software (USEPA, 2012).

RESULTS

In Table – 1, the acute toxicity prediction data were tabulated for both artificial and natural preservatives and these compounds were selected along with their CAS (Chemical Abstracts Services) no. Out of the 16 artificial preservatives, the predicted values of LD_{50} (mg/kg) were obtained by using T.E.S.T. of only 8 compounds viz. benzoic acid (1180.99), ethyl *para*-hydroxybenzoate (2695.96) lactic acid (4723.98), propionic acid (475.28), sorbic acid (1873.12), butylated hydroxytoluene (1639.84), butylated hydroxyanisole (1475.73) and formaldehyde (85.97). Other 8 compounds viz. potassium sorbate, sodium benzoate, calcium benzoate, sodium propionate, disodium EDTA, sodium gallate, sodium nitrite and sodium sulphite unable to predict the LD_{50} values due to CAS no. was not identified by the software.

It was also observed that out of the 16 natural preservatives, the predicted values of LD_{50} (mg/kg) were determined by using T.E.S.T. of 15 compounds viz. caffeic acid (2109.96), cinnamic acid (1817.89), ferulic acid (4742.73), gallic acid (3912.42), oleuropein (4101.86), thymol (714.92), eugenol (2138.45), ascorbic acid (14329.94), tartaric acid (4117.91), malic acid (2600.88), fumaric acid (1000.03), tocopherol acetate (4047.66), carvacrol (1018.17), citral (3022.51) and allin (1502.43). Only 1 compound namely citric acid unable to predict the LD_{50} values due to CAS number was not identified by the software (Table – 1).

All the predicted chemicals for both artificial and natural compounds were very well expressed in the model database by the statistical analysis as per FDA cluster model fit results were tabulated (Table – 1). In this software, the similarity analysis was showed very close similarity with other for related chemicals. The predicted value along with statistically significant value (R^2 and Q^2) for individual chemical was obtained by T.E.S.T. and individual data of chemical from FDA model result for artificial (Fig. 1, 2, 3, 4, 5, 6, 7 and 8) and natural (Fig. 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22 and 23) preservatives were represented graphically.

Sl.	Synthetic preservatives	CAS No.	Predicted	Natural preservatives	CAS No.	Predicted
No.			LD ₅₀	_		LD ₅₀
			(mg/kg)			(mg/kg)
1.	Potassium sorbate	24634-61-5	n.f.	Caffeic acid	331-39-5	2109.96
2.	Sodium benzoate	532-32-1	n.f.	Cinnamic acid	140-10-3	1817.89
3.	Calcium benzoate	2090-05-3	n.f.	Ferulic acid	1135-24-6	4742.73
4.	Benzoic acid	65-85-0	1180.99	Gallic acid	149-91-7	3912.42
5.	Ethyl para- hydroxybenzoate	120-47-8	2695.96	Oleuropein	32619-42-4	4101.86
6.	Lactic acid	50-21-5	4723.98	Thymol	89-83-8	714.92
7.	Sodium propionate	137-40-6	n.f.	Eugenol	97-53-0	2138.45
8.	Propionic acid	79-09-4	475.28	Citric acid	5949-29-1	n.f.
9.	Sorbic acid	110-44-1	1873.12	Ascorbic acid	50-81-7	14329.94
10.	Disodium EDTA	6381-92-6	n.f.	Tartaric acid	87-69-4	4117.91
11.	Butylated hydroxytoluene	128-37-0	1639.84	Malic acid	6915-15-7	2600.88
12.	Butylated hydroxyanisole	25013-16-5	1475.73	Fumaric acid	110-17-8	1000.03
13.	Sodium gallate	2053-21-6	n.f.	Tocopherol acetate	58-95-7	4047.66
14.	Sodium nitrite	7632-00-0	n.f.	Carvacrol	499-75-2	1018.17
15.	Sodium sulphite	7757-83-7	n.f.	Citral	5392-40-5	3022.51
16.	Formaldehyde	50-00-0	85.97	Allin	556-27-4	1502.43

 Table 1. QSAR prediction of acute toxicity (LD₅₀) in rat at oral exposure for synthetic and natural preservatives by using T.E.S.T. software

n.f. = Not found in T.E.S.T.

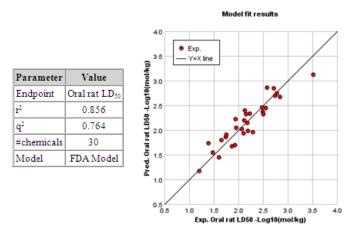


Fig. 1. Statistical analysis and graph for benzoic acid prediction

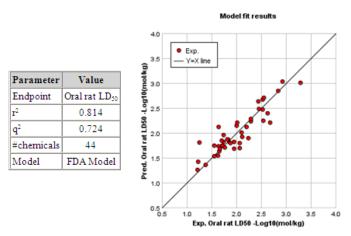


Fig. 2. Statistical analysis and graph for ethyl para-hydroxybenzoate prediction

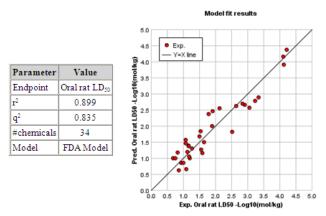


Fig. 3. Statistical analysis and graph for lactic acid prediction

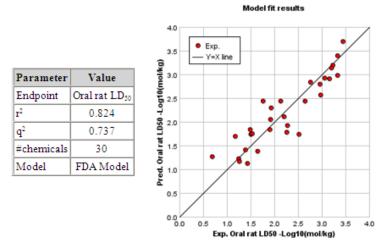


Fig. 4. Statistical analysis and graph for propionic acid prediction

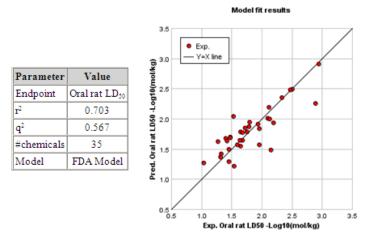


Fig. 5. Statistical analysis and graph for sorbic acid prediction

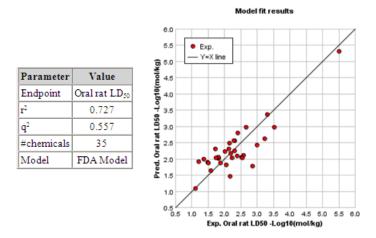


Fig. 6. Statistical analysis and graph for butylated hydroxytoluene prediction

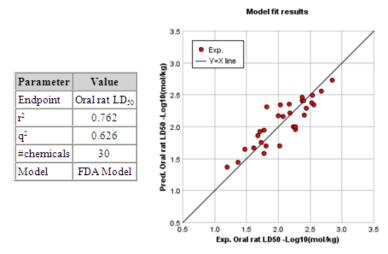


Fig. 7. Statistical analysis and graph for butylated hydroxyanisole prediction

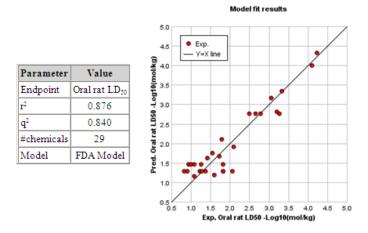


Fig. 8. Statistical analysis and graph for formaldehyde prediction

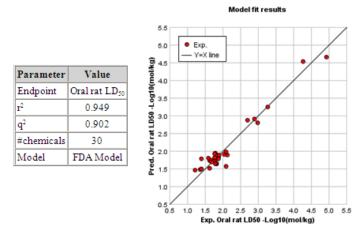


Fig. 9. Statistical analysis and graph for caffeic acid prediction

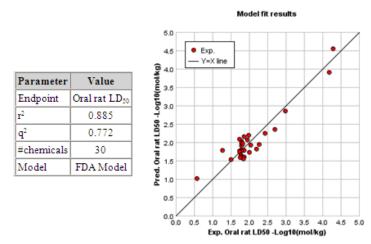


Fig. 10. Statistical analysis and graph for cinnamic acid prediction

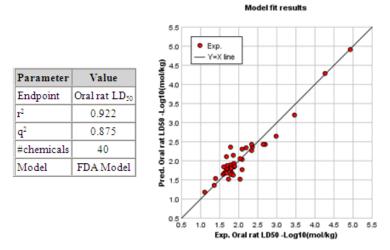


Fig. 11. Statistical analysis and graph for ferulic acid prediction

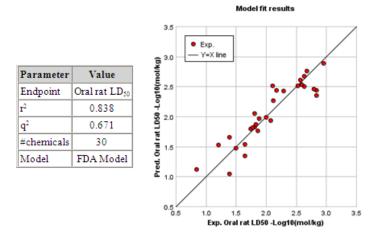


Fig. 12. Statistical analysis and graph for gallic acid prediction

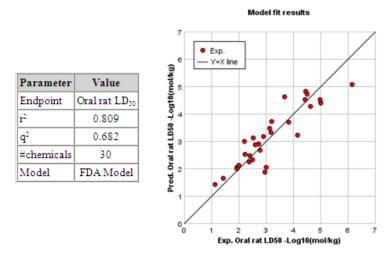


Fig. 13. Statistical analysis and graph for oleuropein prediction

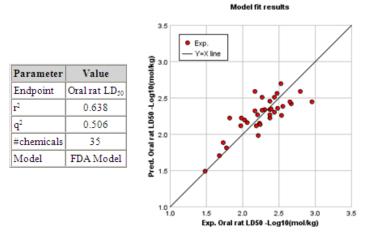


Fig. 14. Statistical analysis and graph for thymol prediction

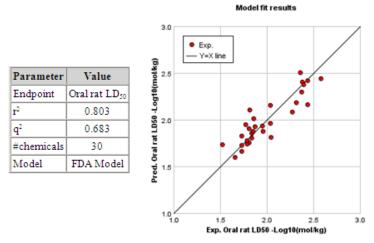


Fig. 15. Statistical analysis and graph for eugenol prediction

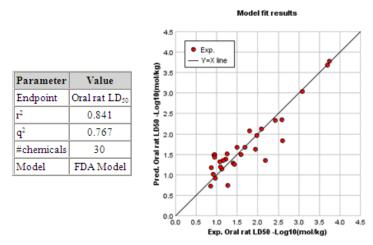


Fig. 16. Statistical analysis and graph for ascorbic acid prediction

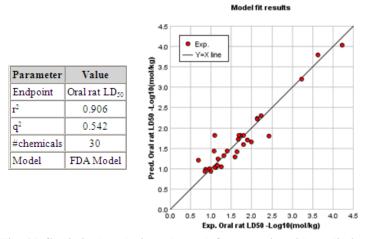


Fig. 17. Statistical analysis and graph for tartaric acid prediction

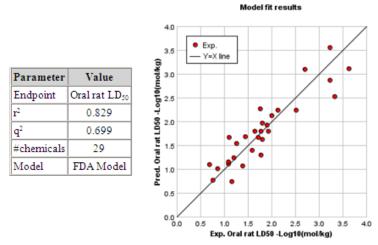


Fig. 18. Statistical analysis and graph for malic acid prediction

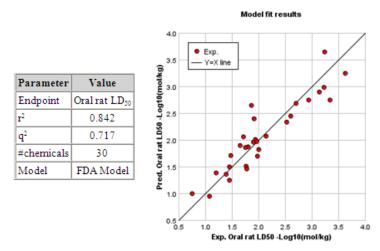


Fig. 19. Statistical analysis and graph for fumaric acid prediction

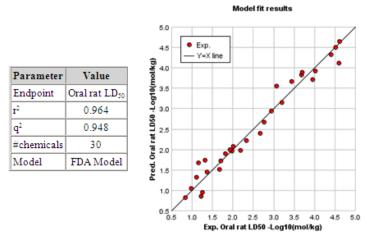


Fig. 20. Statistical analysis and graph for tocopherol acetate prediction

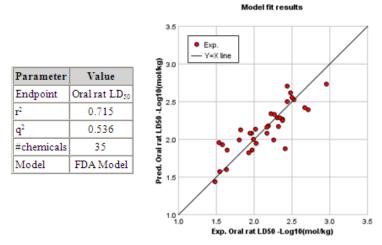


Fig. 21. Statistical analysis and graph for carvacrol prediction

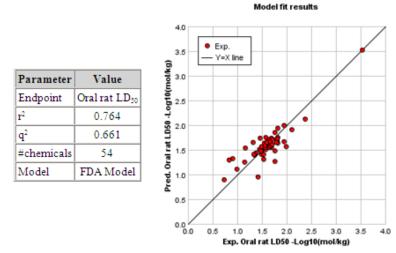


Fig. 22. Statistical analysis and graph for citral prediction

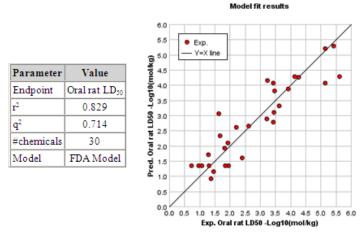


Fig. 23. Statistical analysis and graph for allin prediction

DISCUSSION

The preservation of raw and processed foods always requires the chemical compounds known as preservatives, which protect the foods from bacteria and fungi (Soeda et al., 1966; Close and Nielsen, 1976). The preservatives are also very important to keeping food safe during usage and storage (Faruque, 2012). The present prediction results by using QSAR modeling software indicated that artificial preservatives viz. formaldehyde, and propionic acid and also benzoic acid were predicted lowest LD_{50} value when compared to natural preservatives except thymol, fumaric acid and carvacrol (Table – 1). Moreover, the acute toxicity prediction with special reference to LD_{50} value for both artificial and natural preservatives exposed orally in rat clearly revealed that natural preservatives have higher the LD_{50} value compared to synthetic preservatives. It was well known by many researchers that food preservatives are safer when the chemicals are in natural origin as bioactive compound (Ou and Kwok, 2004; Narasimhan and Dhake, 2006; Chanwitheesuk et al., 2007; Pérez-Díaz and McFeeters, 2010; Mahiwal et al., 2012; Regnier et al., 2012; Belda-Galbis et al., 2013; Mohanka and Priyanka, 2014)

It was documented that artificial food preservatives such as sodium benzoate and sodium nitrite with a high and mid dose of nitrite compared to benzoate showed mortality in offspring of rat (Shuval and Gruener, 1972; Vorhees et al., 1984) while benzoic acid revealed acute toxicity with clinical symptoms in rats (Bio-Fax, 1973). It was reported few works on QSAR modeling of individual compound used as preservatives but the present findings were elaborative

study on acute toxicity prediction in rat after oral exposure of artificial and natural preservatives by using T.E.S.T. QSAR modeling software.

QSAR technique can be beneficial for discovering the relationship between the chemical molecular structures and their acute toxicities. It was known that the T.E.S.T. software estimates the predicted rat oral LD_{50} values by calculating several 2D molecular descriptors (USEPA, 2012). These molecular descriptors have used to detect toxicity prediction in mammals by QSAR modeling (Gombar and Jain, 1987; Gombar and Enslein, 1990; Hall et al., 1991; Ruiz et al., 2012). Linear regression analysis is widely used as a final statistical method at the end of QSAR study (Xu, 2004; Li et al., 2010).

In this present study, the rat oral LD_{50} predicted values were obtained for QSAR modeling by using T.E.S.T. software. It was reported that T.E.S.T. software has better performance than other softwares like ADMET and TOPKAT for the prediction of sulfur mustard and its breakdown products on mammal (Ruiz et al., 2012). Previously worked on the acute toxicity prediction of fluroqunilone antibiotics in rat with T.E.S.T. software also predicted well with exact LD_{50} value as per R² value of individual compound (Talapatra et al., 2015). According to studies by Canadian Center for Occupational Health & Safety (2012) and Ruiz et al. (2012), the toxicity ranges were determined as super toxic (<5 mg/kg), extremely toxic (5–50 mg/kg), very toxic (50–500 mg/kg), moderately toxic (500–5,000 mg/kg), slightly toxic (5,000–15,000 mg/kg) and practically non-toxic (>15,000 mg/kg) in the previous study of chemicals. The food preservatives were selected in the present study as per usage (Health Canada, 2012) but formaldehyde is non-permitted food preservatives (Janny, 2010). The present predicted acute toxicity results with special reference to LD_{50} values of artificial and natural preservatives were showed that artificial preservatives viz. formaldehyde, and propionic acid and also benzoic acid were predicted lowest LD_{50} value when compared to natural preservatives except thymol, fumaric acid and carvacrol. It is suggested natural preservatives might be used after conducting bioassay to correlate the present toxicity data from raw and processed food management perspective.

CONCLUSION

The acute toxicity with special reference to LD_{50} values in rat after oral exposure for 16 artificial and natural preservatives. Among 16 artificial preservatives, only 8 compounds were predicted and 15 compounds were predicted for artificial and natural compounds respectively. The results suggested that predicted acute toxicity of 3 artificial preservatives viz. formaldehyde, and propionic acid (>50 and <500 mg/kg) as very toxic and also benzoic acid (>500 and <500 mg/kg) as moderately toxic were predicted lowest LD_{50} value when compared to natural preservatives were within range of >500 and <5000 mg/kg as moderately toxic. This software has a potent capability to predict rat oral LD_{50} value with suitable inbuilt programming of QSAR modeling with 2D molecular descriptors and similar test chemicals by calculating test sets and training set (Ruiz et al., 2012; USEPA, 2012). Although the benzoic acid and its derivatives were reported moderately toxic in experimental study as well as QSAR modeling (WHO, 2000; Li et al., 2010). Furthermore, natural preservatives in food are safer than artificial chemicals (Regnier et al., 2012) and management practices are suggested in relation to innovation of new brands of natural preservatives can be used from plant origin (Pérez-Díaz and McFeeters, 2010; Mohanka and Priyanka, 2014). This present prediction work was based on only single QSAR modeling approach but should need further investigation by using other softwares.

REFERENCES

Belda-Galbis, C.M., Leufvén, A., Martínez, A. and Rodrigo, D. (2013): Quantitative assessment of citral antimicrobial potential at different temperatures. In: Microbial pathogens and strategies for combating them: science, technology and education (ed. Mendez-Vilas, A.). Formatex 1257-1264.

Bhullar, K.S., Jha, A., Youssef, D. and Vasantha Rupasinghe, H.P. (2013): Curcumin and its carbocyclic analogs: Structure-activity in relation to antioxidant and selected biological properties. Molecules 18: 5389-5404.

Bio-Fax, (1973): Benzoic acid. Industrial Bio-Test Laboratories, Inc., Northbrook, IL.

CAL (Consumers, Association Limited) (1988): Additives. Saunder, B. and Bradley, A. (eds). Hazel and Viney Ltd. British Library, Great Britain pp. 43, 131-139.

Canadian Center for Occupational Health & Safety, (2012): What is an LD_{50} and LC_{50} . Available online: http://www.ccohs.ca/oshanswers/chemicals/LD50.html#_1_6 (accessed on 26 July).

Cassano A., Manganaro A., Martin T., Young, D., Piclin, N., Pintore, M., Bigoni, D. and Benfenati, E. (2010): CAESAR models for developmental toxicity. Chemistry Central Journal 4(Suppl 1): S4 (doi:10.1186/1752-153X-4-

S1-S4).

Chanwitheesuk, A., Teerawutgulrag, A., Kilburn, J.D. and Rakariyatham, N. (2007): Antimicrobial gallic acid from *Caesalpinia mimosoides* Lamk. Food Chem. 100: 1044-1048.

Chapman, J.S. (1998): Characterizing bacterial resistance to preservatives and disinfectants. Int. Biodeter. Biodegr. 41: 241-45.

Chaudhary, J., Rajpal, A.K., Judge, V., Narang, R. and Narasimhan, B. (2008): Synthesis, antimicrobial evaluation and QSAR analysis of caprylic acid derivatives. Scientia Pharm. 76.

Choplin, F. (2005): Comprehensive medicinal chemistry, Corwin Hansch, Vol 4, Elsevier Pergamon, Oxford, 33-57. Close, J. and Nielsen, P.A. (1976): Resistance of a strain of *Pseudomonas capacia* to ester of p-hydroxybenzoic acid. Appl. Enviorn. Microbiol. 31: 718-22.

Cutler, H.G. (1995): Natural flavor compounds as potential antimicrobials insecticides and medicinal. Agro food Ind. Hi-Tech 6: 19-23.

Erasto, P., Bojase-Moleta, G. and Majinda, R.R.T. (2004): Antimicrobial and antioxidant flavonoids from the root wood of *Bolusanthus speciousus*. Phytochemistry 65: 875-880.

Faruque, S.M. (ed) (2012): Food borne and waterborne bacterial pathogens: Epidemiology, Evolution and Molecular Biology. Caister Academic Press. ISBN 978-1-908230-06-5.

Golbraikh, A. and Tropsha, A. (2002): Beware of q²! J. Mol. Graph. Model. 20: 269-276.

Golbraikh, A., Shen, M., Xiao, Z., Xiao, Y.-D., Lee, K.-H. and Tropsha, A. (2003): Rational selection of training and test sets for the development of validated QSAR models. J. Computer-Aided Mol. Design 17: 241-253.

Gombar, V.K. and Enslein, K. (1990): Quantitative structure-activity relationship (QSAR) studies using electronic descriptors calculated from topological and molecular orbital (MO) methods. QSAR 9: 321-325.

Gombar, V.K. and Jain, D.V.S. (1987): Quantification of molecular shape and its correlation with physico-chemical properties. Indian J. Chem. 24A: 554-555.

Gramatica, P. and Pilutti, P. (2004): Evaluation of different statistical approaches for the validation of quantitative structure-activity relationships. Ispra, Italy: The European Commission - Joint Research Centre, Institute for Health & Consumer Protection - ECVAM.

Hall, L.H., Mohney, B. and Kier, L.B. (1991): The electrotopological state: Structure information at the atomic level for molecular graphs. J. Chem. Inf. Comput. Sci. 31: 76-82.

Health Canada, (2012): Transition guide: Understanding and using the lists of permitted food additives. November. http://www.hc-sc.gc.ca/fn-an/securit/addit/list/transition-eng.php

Janny, M.A. (2010): Formaldehyde in noodlefish. Food Safety Focus. Issue 42 (http://www.cfs.gov.hk/ english/multimedia/multimedia_pub_pub_fsf_42_01.html).

Li, Z., Sun, Y., Yan, X. and Meng, F. (2010): Study on QSTR of benzoic acid compounds with MCI. Int. J. Mol. Sci. 11: 1228-1235.

Mahiwal, K., Kumar, P. and Narasimhan, B. (2012): Synthesis, antimicrobial evaluation, ot-QSAR and mt-QSAR studies of 2-amino benzoic acid derivatives. Med. Chem. Res. 21(3): 293-307.

Martin, T.M., Harten, P., Venkatapathy, R., Das, S. And Young, D.M. (2008): A hierarchical clustering methodology for the estimation of toxicity. Toxicol. Mech. Methods 18: 251-266.

Mohanka, R. and Priyanka (2014): Plant extract as natural food preservative against spoilage fungi from processed food. Int. J. Curr. Microbiol. App. Sci. 3(8): 91-98.

Narasimhan, B., Belasare, D., Pharande, D., Mourya, V. and Dhake, A. (2004): Esters, amides and substituted derivatives of cinnamic acid: synthesis, antimicrobial activity and QSAR investigations. Eur. J. Med. Chem. 39: 827-34.

Narasimhan, B. and Dhake, A.S. (2006): Antibacterial constituents from nut shell of *Anacardium occidentale*. Planta Indica 2(2): 4-7.

Narasimhan, B., Kothawade, U.R., Pharande, D.S., Mourya, V.K. and Dhake, A.S. (2003): Syntheses and QSAR studies of sorbic, cinnamic and ricinoleic acid derivatives as potential antibacterial agents. Indian J. Chem. 42(B): 2828-34.

Narasimhan, B., Mourya, V.K. and Dhake A.S. (2006): Design, synthesis, antibacterial and QSAR studies of myristic acid derivatives. Bioorg. Med. Chem. Lett. 6: 3023-29.

Narasimhan, B., Ohlan, S., Ohlan, R., Judge, V. and Narang, R. (2009): Hansch analysis of veratric acid derivatives as antimicrobial agents. Eur. J. Med. Chem. 44(2): 689-700.

Netzeva, T., Worth, A.P., Aldenberg, T., Benigni, R., Cronin, M.T., Gramatica, P., Jaworska, J., Kahn, S., Klopman, G., Marchant, C.A., Myatt, G., Nikolova-Jeliazkova, N., Patlewicz, G., Perkins, R., Roberts, D., Schultz, T., Stanton, D., van de Sandt, J., Tong, W., Veith, G. and Yang, C. (2005): Current status of methods for defining the

applicability domain of (quantitative) structure-activity relationships. The report and recommendations of ECVAM Workshop 52, Altern. Lab. Anim. 33: 1-19.

Ou, S. and Kwok, K.C. (2004): Ferulic acid: pharmaceutical functions, preparation and applications in foods. J. Sci. Food Agri. 84: 1261-1269.

Pallas 3.1.1.2, ADME-Tox software (2000): Computer Drug International Inc., U.S.A.

Patil, P.A., Amnerkar, N.D., Sandeep, S., Pathare, S.S. and Bhusari, K.P. (2014): 3D-QSAR study, synthesis and biological evaluation of p-hydroxy benzohydrazide derivatives as antimicrobial agents. Der Pharma Chemica 6 (6): 300-312.

Pérez-Díaz, I.M. and McFeeters, R.F. (2010): Preservation of acidified cucumbers with a natural preservative combination of fumaric acid and allyl isothiocyanate that target lactic acid bacteria and yeasts. J. Food Sci. 75: M204-M208.

Puupponen, P.R., Nohynek, L., Meier, C., Kahkonen, M.M., Heinonen, A. and Hopia, K.M. (2001): Antimicrobial properties of phenolic compounds from Berries. J. Applied Microboil. 90: 494-507.

Regnier, T., Combrinck, S. and Du Plooy, W. (2012): Essential oils and other plant extracts as food preservatives, In: Progress in Food Preservation (Bhat, R., Alias, A.K. and Paliyath, G. eds.), Wiley-Blackwell, Oxford, UK (doi: 10.1002/9781119962045) Ch 26.

Roy, P.P., Kovarich, S. and Gramatica, P. (2011): QSAR model reproducibility and applicability: A case study of rate constants of hydroxyl radical reaction models applied to polybrominated diphenyl ethers and (benzo-)triazoles. J. Comput. Chem. 32: 2386-2396.

Ruiz, P., Begluitti, G., Tincher, T., Wheeler, J. and Mumtaz, M. (2012): Prediction of acute mammalian toxicity using QSAR methods: A case study of sulfur mustard and its breakdown products. Molecules 17: 8982-9001.

Sarova, D., Kapoor, A., Narang, R., Judge, V. and Narasimhan, B. (2011): Dodecanoic acid derivatives: Synthesis, antimicrobial evaluation and development of one-target and multi-target QSAR models. Med. Chem. Res. 20(6): 769-81.

Schultz, T.W., Hewitt, M., Netzeva, T.I. and Cronin, M.T.D. (2007): Assessing applicability domains of toxicological QSARs: Definition, confidence in predicted values, and the role of mechanisms of action. QSAR Comb. Sci. 26: 238-254.

Shuval, H.L. and Gruener, N. (1972): Epidemiological and toxicological aspects of nitrates and nitrites in the environment. Ninety-ninth Annual Meeting of the American Public Health Association. Minneapolis, Minnesota.

Soeda, M., Otomo, M., Ome, M. and Kawashima, K. (1966): Studies on anti-bacterial and anti-fungal activity of Cape Aloe. Nippon Saikingaku Zasshi 21: 609-614.

Souza, E.L.D., Lima, T.L.M.S.O., Tarjano, V.N. and Filho, J.M.B. (2005): Antimicrobial effectiveness of spices an approach for use in food conservation systems. Braz. Arch. Biol. Tech. 48: 1-13.

Talapatra, S. N., Misra, D., Banerjee, K., Banerjee, P. and Swarnakar, S. (2015): QSAR modeling for acute toxicity prediction of fluroquinolone antibiotics by using software. Int. J. Adv. Res. 3 (6): 225-240.

Talete (2006): Dragon Version 5.4 (http://www.talete.mi.it/dragon_net.htm).

Tetko, I.V., Sushko, I., Pandey, A.K., Zhu, H., Tropsha, A., Papa, E., Oberg, T., Todeschini, R., Fourches, D. and Varnek, A. (2008): Critical assessment of QSAR models of environmental toxicity against *Tetrahymena pyriformis*: Focusing on applicability domain and overfitting by variable selection. J. Chem. Inf. Model. 48: 1733-1746.

TOPKAT (2004): User Guide Version 6.2 Accelrys: San Diego, CA, USA.

Tropsha, A. and Golbraikh, A. (2007): Predictive QSAR modeling workflow, model applicability domains, and virtual screening. Curr. Pharm. Des. 13: 3494-3504.

USEPA (United States Environmental Protection Agency) (2012): T. E. S. T Tool, User's Guide for T.E.S.T, Version 4.1.; A Program to Estimate Toxicity from Molecular Structure, Cincinatti, OH, USA.

User Manual ADMET, Version 5.5 (2011): Simulation Plus Inc, S. P.: Lancaster, CA, USA.

Valentina, P., Lango, K. and Engels, M. (2009): Rationalization of physicochemical characters of 2-Phenyl-3hydroxy-4(1H)-quinolinone-7-carboxylic acid analogs as topoisomerase inhibitors: A QSAR approach. Indian J. Pharm. Educ. Res. 43(3): 284-289.

Venkataramana, C.H.S., Ramya Sravani, K.M., Singh, S.S. and Madhavan, V. (2011): In-silico ADME and toxicity studies of some novel indole derivatives. J. Appl. Pharm. Sci. 1 (10): 159-162.

Vorhees, C.V., Butcher, R.E., Brunner, R.L. and Wooten, V. (1984): Developmental toxicity and psychotoxicity of sodium nitrite in rats. Food Chem. Toxic. 22(1): 1-6.

WHO (World Health Organization) (2000): Benzoic acid and sodium benzoate. International Programme on Chemical Safety. Concise International Chemical Assessment Document no. 26.

Worth, A.P., Bassan, A., DeBruijn, J., Gallegos Saliner, A., Netzeva, T., Patlewicz, G., Pavan, M., Tsakovska, I. and Eisenreich, S. (2007): The role of the European Chemicals Bureau in promoting the regulatory use of (Q)SAR methods. SAR and QSAR Env. Res. 18 (1-2): 111-125.

Xu, S.J. (2004): Computer-assisted drug molecular design. Chemical Industry Press, Beijing, China.

Zhu, H., Martin, T.M., Ye, L., Sedykh, A., Young, D.M. and Tropsha, A. (2009): Quantitative structure-activity relationship modeling of rat acute toxicity by oral exposure. Chem. Res. Toxicol. 22: 1913-1921.