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**RESEARCH ARTICLE**

**GC-MS ANALYSIS AND ANTIMICROBIAL ACTIVITY OF SUDANESE *CYPERUS ESCULENTUS*
 (CYPERACEAE) FIXED OIL.**

Abdel Karim M^{1,*} and Fath El-Rahman A².

1. Sudan University of Science and Technology, Faculty of Science, Dept. of Chemistry.
2. Omdurman Islamic University, Faculty of Education, Dept. of Chemistry.

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Abstract

The present study was designed to investigate the chemical constituents of *Cyperus esculentus* seed oil and to evaluate its potential antimicrobial activity. 21 components were detected by GC-MS analysis. Major constituents are: 9-octadecenoic acid (46.24%), hexadecanoic acid (19.27%), 9,12-octadecadienoic acid (13.62%), and methyl stearate (10.88%). Butylated hydroxytoluene, a potent antioxidant, was detected as a minor constituent (0.10%). The antibacterial activity of the oil was evaluated via cup plate agar diffusion assay against six standard human pathogens (Gram positive: *Staphylococcus aureus* and *Bacillus subtilis*; Gram negative: *Escherichia coli* and *Pseudomonas aeruginosa* and the fungi *Candida albicans* and *Aspergillus niger*). The oil showed different antimicrobial responses against test organisms. It gave significant activity against the fungus *Candida albicans* and partial activity against *Staphylococcus aureus*. It seems that the oil is a lead for further optimization.

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Introduction:-

Cyperus esculentus is a grass-like plant of the family Cyperaceae (Sedge family) which is widely distributed in many north temperature locations within south Europe as its probable origin (Mason, 2005). Tiger nut (*Cyperus esculentus*) is not really a nut but a small tuber, first discovered some 4000 years ago in ancient Egypt and is cultivated today in China, Spain and West Africa. *Cyperus esculentus* "tiger nut" is also known as chufa, earth nut, yellow nut sedge, groundnut, rush nut, and edible galingale (Oderinde and Tairu, 1988). The plant develops as a series of shoots, bulbs and stem tubers connected by brown wiry rhizomes which are strengthened by lignification of the inner cortex. Tubers are small, 1-2 cm in diameter, and are borne at intervals along the rhizomes. (Garg, 1967).

Along with a high-energy content (starch, fats, sugars and proteins), the plant is rich in minerals such as phosphorous and potassium and in vitamins E, C, soluble glucose and oleic acid. Typically, 100g tiger nuts contain 386 kcal (1635 kJ) as: 7% proteins, 26% fats (oils), 31% starch, 21% glucose. It contains 26% fibre of which 14% is non-soluble. *Cyperus esculentus* underground tubers are considered as an important food for waterfowl (McAfee, 1939) and sand hill cranes on wintering areas (Guthery, 1976).

Cyperus esculentus, an underutilized crop, was reported to be high in dietary fibre content, which could be effective in the treatment and prevention of many diseases including colon cancer, coronary heart diseases, obesity, diabetes

Corresponding Author:- Abdel Karim M.

Address:- Sudan University of Science and Technology, Faculty of Science, Dept. of Chemistry.

and gastrointestinal disorders (Anderson *et.al.* 1994). Tiger nut has also been reported to be used in the treatment of flatulence, indigestion, diarrhea, dysentery, and excessive thirst (Chevallier, 1996). They are also used in Brazil for the treatment fever (De Abreu Matos, 2008) and in losing weight (Borges *et.al.*, 2008).

Materials and Methods:-

Plant material:-

Tubers of *Cyperus esculentus* were purchased from the local market – Omdurman, Sudan. The plant was kindly authenticated by Institute of Aromatic and Medicinal Plants- Khartoum, Sudan.

Instruments:-

A Shimadzo GC-MS-QP2010 Ultra instrument with a RTX-5MS column (30m, length ; 0.25mm diameter ; 0.25 μ m, thickness) was used for GC-MS analysis .

Test organisms:-

Cyperus esculentus oil was screened for antibacterial and antifungal activities using the standard microorganisms shown in Table(1).

Table 1:- Test organisms.

Ser. No	Microorganism	Type
1	<i>Bacillus subtilis</i>	G+ve
2	<i>Staphylococcus aureus</i>	G+ve
3	<i>Pseudomonas aeruginosa</i>	G-ve
4	<i>Escherichia coli</i>	G-ve
5	<i>Aspergillus niger</i>	fungi
6	<i>Candida albicans</i>	fungi

Methods:-

Extraction of oil from *Cyperus esculentus*

Dry-powdered tubers of *Cyperus esculentus* (200g) were macerated with n-hexane at room temperature for 48h..The solvent was removed under reduced pressure and the oil was kept in the fridge at 4°C for further manipulation.

Esterification of oil:-

A Methanolic solution of sodium hydroxide was prepared by dissolving (2g) of sodium hydroxide in 100ml methanol. A stock solution of methanolic sulphuric acid was prepared by mixing (1ml)of concentrated sulphuric acid with (99ml) methanol.

The oil(2ml) was placed in a test tube and (7ml) of alcoholic sodium hydroxide were added followed by (7ml) of alcoholic sulphuric acid. The tube was stoppered and shaken vigorously for five minutes and then left overnight.(2ml) of supersaturated sodium chloride were added, then (2ml) of n- hexane were added and the tube was vigorously shaken for five minutes .The hexane layer was then separated.(5 μ l) of the hexane extract were mixed with 5ml diethyl ether . The solution was filtered and the filtrate(1 μ l) was injected in the GC-MS vial.

GC-MS analysis:-

Cyperus esculentus fixed oil was analyzed by gas chromatography – mass spectrometry. A Shimadzo GC-MS-QP2010 Ultra instrument with a RTX-5MS column (30m, length ; 0.25mm diameter ; 0.25 μ m, thickness)was used. Helium (purity; 99.99 %) was used as carrier gas. Oven temperature program is given in Table 2, while other chromatographic conditions are depicted in Table 3.

Table 2:- Oven temperature program

Rate	Temperature(C)	Hold time (min. ⁻¹)
-	60.0	0.00
10.00	300.0	0.00

Table 3:- Chromatographic conditions

Column oven temperature	1300.0 °C
Injection temperature	280.0 °C
Injection mode	Split
Flow control mode	Linear velocity
Pressure	93.1KPa
Total flow	50.0ml/ min
Column flow	1.50ml/sec
Linear velocity.	44.7cm/sec
Purge flow	3.0ml/min.
Spilt ratio	- 1.0

Antimicrobial assay:-**Preparation of bacterial suspensions:-**

One ml aliquots of 24 hours broth culture of the test organisms were aseptically distributed onto nutrient agar slopes and incubated at 37°C for 24 hours. The bacterial growth was harvested and washed off with sterile normal saline, and finally suspended in (100 ml) of normal saline to produce a suspension containing about 10⁸-10⁹ colony forming units per ml. The suspension was stored in the refrigerator at 4°C until used. The average number of viable organism per ml of the stock suspension was determined by means of the surface viable counting technique.

Serial dilutions of the stock suspension were made in sterile normal saline in tubes and one drop volumes (0.02 ml) of the appropriate dilutions were transferred by adjustable volume micropipette onto the surface of dried nutrient agar plates. The plates were allowed to stand for two hours at room temperature for the drop to dry, and then incubated at 37°C for 24 hours.

Preparation of fungal suspensions:-

Fungal cultures were maintained on dextrose agar incubated at 25°C for four days. The fungal growth was harvested and washed with sterile normal saline, and the suspension was stored in the refrigerator until used.

Testing for antibacterial activity:-

The cup-plate agar diffusion method was adopted, with some minor modifications, to assess the antibacterial activity. (2ml) of the standardized bacterial stock suspension were mixed with (200 ml) of sterile molten nutrient agar which was maintained at 45°C in a water bath. (20 ml) Aliquots of the incubated nutrient agar were distributed into sterile Petri dishes. The agar was left to settle and in each of these plates which were divided into two halves, two cups in each half (10 mm in diameter) were cut using sterile cork borer (No 4), each one of the halves was designed for one of the test solutions. Separate Petri dishes were designed for standard antibacterial chemotherapeutics (ampicillin and gentamycin).

The agar discs were removed, alternate cups were filled with(0.1 ml) samples of each test solution using adjustable volume microtiter pipette and allowed to diffuse at room temperature for two hours. The plates were then incubated in the upright position at 37°C for 24 hours.

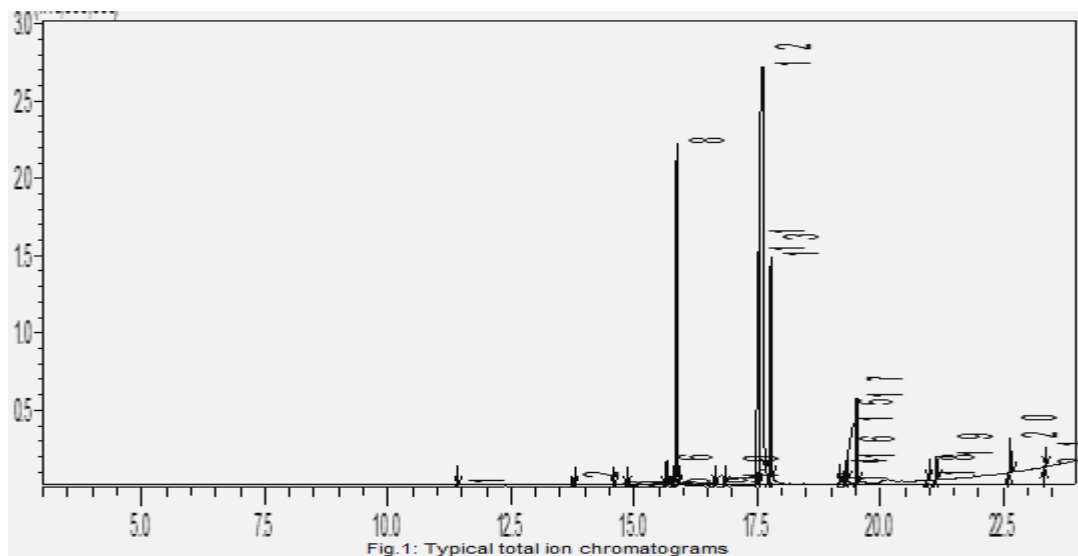
The above procedure was repeated for different concentrations of the test solutions and the standard chemotherapeutics. After incubation, the diameters of the resultant growth inhibition zones were measured in triplicates and averaged.

Results and Discussion:-**GC-MS analysis of *Cyperus esculentus* fixed oil:-**

GC-MS analysis of *Cyperus esculentus* oil was conducted and the constituents were identified by comparison with the MS library (NIST) and further confirmed by interpreting the observed fragmentation pattern. Comparison with the database on MS library gave about 90-95% match.

Constituents of oil:-

The GC-MS spectrum of the studied oil revealed the presence of 21 components (Table 4). The typical total ion chromatograms (TIC) is displayed in Fig.1.

Table 4:- Constituents of *Cyperus esculentus* oil

Peak#	R.Time	Area	Area%	Name
1	11.396	276131	0.10	Butylated Hydroxytoluene
2	13.747	1040707	0.38	Methyl tetradecanoate
3	14.559	25275	0.01	6-Octadecenoic acid, methyl ester
4	14.822	93781	0.03	Pentadecanoic acid, methyl ester
5	15.614	153749	0.06	Methyl hexadec-9-enoate
6	15.658	2544921	0.92	9-Hexadecenoic acid, methyl ester, (Z)-
7	15.751	94133	0.03	cis-10-Nonadecenoic acid, methyl ester
8	15.868	53170840	19.27	Hexadecanoic acid, methyl ester
9	16.621	388831	0.14	cis-10-Heptadecenoic acid, methyl ester
10	16.827	726176	0.26	Heptadecanoic acid, methyl ester
11	17.518	37587908	13.62	9,12-Octadecadienoic acid (Z,Z)-, methyl e
12	17.613	127589980	46.24	9-Octadecenoic acid (Z)-, methyl ester
13	17.776	30023371	10.88	Methyl stearate
14	19.159	161246	0.06	10,13-Eicosadienoic acid, methyl ester
15	19.282	1688801	0.61	Oxiraneoctanoic acid, 3-octyl-, methyl ester
16	19.322	2190882	0.79	11-Eicosenoic acid, methyl ester
17	19.519	10162898	3.68	Methyl 18-methylnonadecanoate
18	20.963	342486	0.12	13-Docosenoic acid, methyl ester
19	21.137	2625595	0.95	Methyl 20-methyl-heneicosanoate
20	22.639	4693698	1.70	Tetracosanoic acid, methyl ester
21	23.353	326153	0.12	Methyl 23-methyl-tetracosanoate
		275907562	100.00	

Some important constituents are discussed below:

9-Octadecenoic acid methyl ester(46.24%):-

The peak at m/z 296, which appeared at R.T. 17.613(Fig.2) in total ion chromatogram, corresponds to $M^+[C_{19}H_{36}O_2]^+$, while the peak at m/z 266 accounts for loss of a methoxy group.

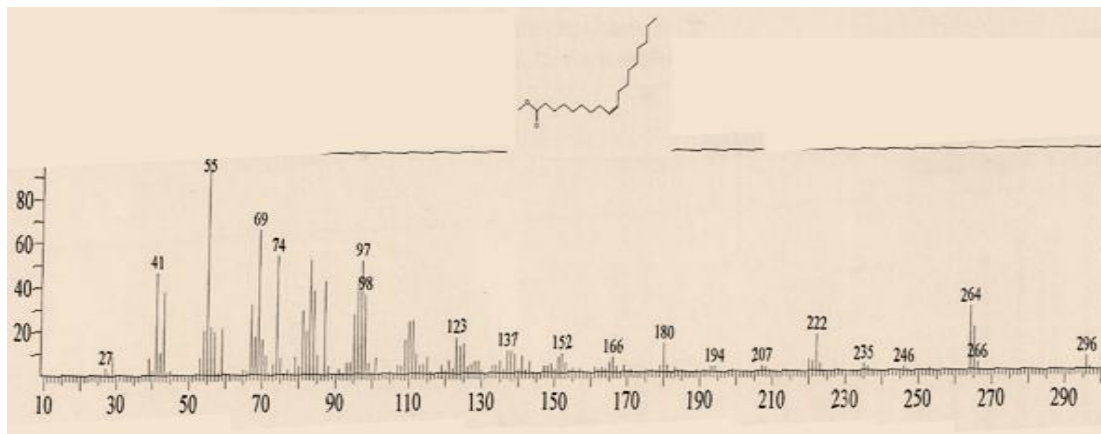


Fig. 2:- Mass spectrum of 9-octadecenoic acid methyl ester.

Oleic acid (9-octadecenoic acid) acid is very common in human diet. The hypotensive potential of olive oil is probably due to this acid (Terese *et.al.*,2008). This acid finds some applications in soap industry and it is used in small amounts in some pharmaceutical products. It is also used as soldening flux in stained glass work. Oleic acid is employed as emollient(Currasco,2002). It is claimed that the consumption of oleate in olive oil has been associated with decreased risk of breast cancer(Martin *et.al.*,1994)

9,12-Octadecadienoic acid methyl ester (13.62%):-

Fig. 3 shows the EI mass spectrum of 9,12-octadecadienoic acid methyl ester. The peak at m/z 294, which appeared at R.T. 17.518 in total ion chromatogram, corresponds to $M^+[C_{19}H_{34}O_2]^+$. The peak at m/z 263 corresponds to loss of a methoxy function.

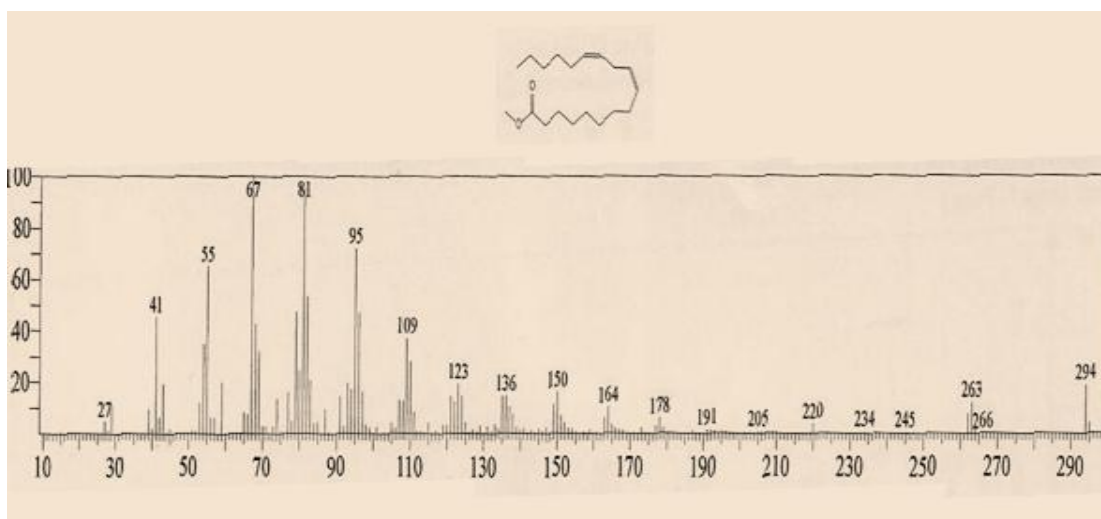


Fig. 3:- Mass spectrum of 9,12-octadecadienoic acid methyl ester.

9,12-Octadecadienoic acid is an essential fatty acid that can not be synthesized by humans and is available through diet(Burr *et.al.*,1930).It belongs to one of the two families of essential fatty acids. It occurs in lipids of cell membrane and is used in the biosynthesis of arachidonic acid . It is converted enzymatically into mono-hydroxy

products which are subsequently oxidized by some enzymes to keto metabolites. Deficiency of linolate caused hair loss and poor wound healing in model animals (Cunnane and Anderson, 1997; Ruthig and Mecklung-Gill, 1999).

Hexadecanoic acid methyl ester (19.27%):-

Mass spectrum of hexadecanoic acid methyl ester is depicted in Fig. 4. The peak at m/z 270, which appeared at R.T. 15.868 corresponds to $M^+[C_{17}H_{34}O_2]^+$ while the peak at m/z 239 is attributed to loss of a methoxyl group.

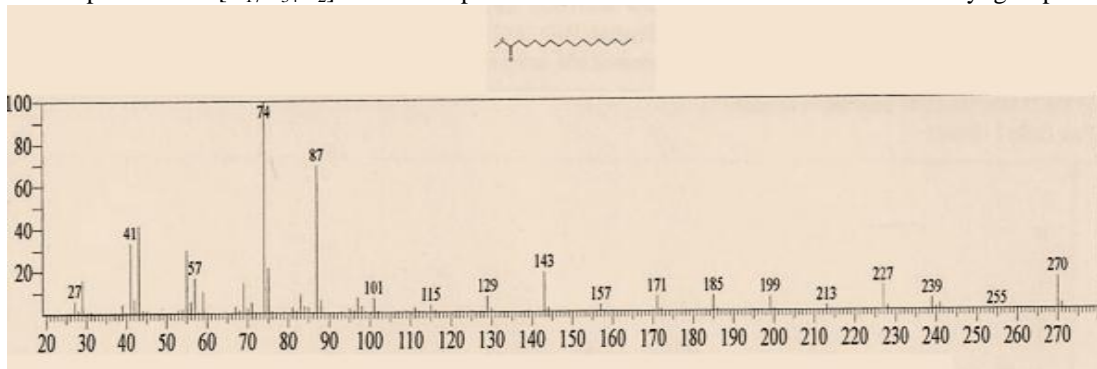


Fig. 4:- Mass spectrum of hexadecanoic acid methyl ester.

Hexadecanoic acid (palmitic acid) is a saturated fatty acid which is very common in plants. It is produced first during the synthesis of fatty acids (Gunstone et al., 2007) and is considered as precursor of long-chain fatty acids. Palmitic acid is a major lipid component of human breast milk (Kingsbury et al., 1961; Jensen et al., 1978). The acid finds applications in soaps and cosmetics industries. It is also used in food industry.

Methyl stearate (10.88%):-

Mass spectrum of methyl stearate is shown in Fig. 5. The peak at m/z 298, which appeared at R.T. 17.776 corresponds to $M^+[C_{19}H_{38}O_2]^+$. The peak at m/z 267 corresponds to loss of a methoxyl function.

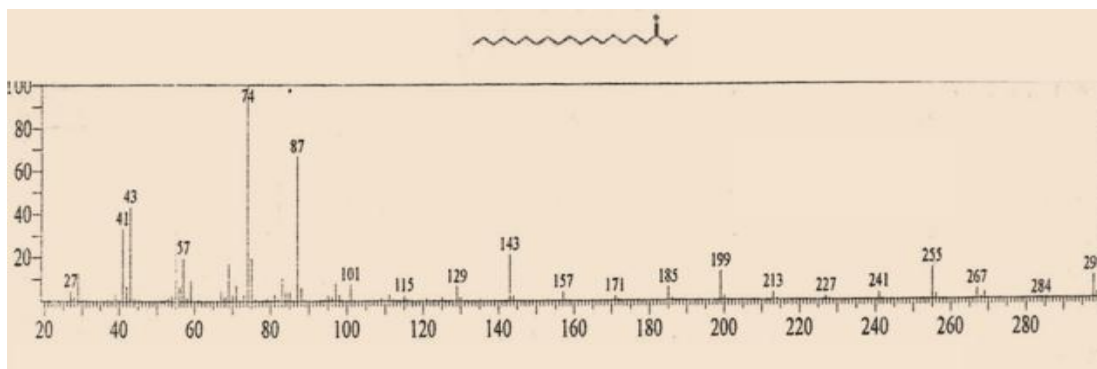


Fig. 5:- Mass spectrum of methyl stearate.

Antibacterial activity:-

In cup plate agar diffusion assay, the oil was screened for antimicrobial activity against six standard human pathogens. The average of the diameters of the growth of inhibition zones are depicted in Table (5). The results were interpreted in commonly used terms (<9mm: inactive; 9-12mm: partially active; 13-18mm: active; >18mm: very active). Tables (6) and (7) represent the antimicrobial activity of standard antibacterial and antifungal chemotherapeutic agents against standard bacteria and fungi respectively.

Table 5:- Antibacterial activity of *Cyperus esculentus* oil

Type	Conc.(mg/ml)	Sa	Bs	Ec	Ps	Ca	An
Oil	100	12	8	-	-	19	8

Table 6:- Antibacterial activity of standard chemotherapeutic agents

Drug	Conc.(mg/ml)	Bs	Sa	Ec	Ps
Ampicilin	40	15	30	-	-
	20	14	25	-	-
	10	11	15	-	-
Gentamycin	40	25	19	22	21
	20	22	18	18	15
	10	17	14	15	12

Table 7:- Antifungal activity of standard chemotherapeutic agent

Drug	Conc.(mg/ml)	An	Ca
Clotrimazole	30	22	38
	15	17	31
	7.5	16	29

Sa.: *Staphylococcus aureus*

Ec.: *Escherichia coli*

Pa.: *Pseudomonas aeruginosa*

An.: *Aspergillus niger*

Ca.: *Candida albicans*

Bs.: *Bacillus subtilis*

The oil was partially active against *Staphylococcus aureus*, but it exhibited significant activity against the fungus: *Candida albicans*.

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