

RESEARCH ARTICLE

AN OVERVIEW OF BUFFALO MILK QUALITY FROM BEED DISTRICT

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..... Manuscript Info Abstract Manuscript History This study was carried out to determine the chemical and Received: 05 May 2022 microbiological quality of buffalo milk. A total of 120 buffalo raw Final Accepted: 08 June 2022 milk samples were collected monthly from beed district, throughout the Published: July 2022 year for this study. In the chemical analysis of buffalo milk samples the mean total solid value, non-solid fat, lipid, protein, lactose, ash and pH Key words:values were detected as 16.38%, 8.56%, 7.04%, 4.36%, 4.19%, 0.72% Buffalo Milk, Chemical Ouality. and 6.55, respectively. Total bacteria count (TCA), coliform, lactic acid Microbiological Quality bacteria (LAB), Escherichia coli, Staphylococcus aureus and yeastmold (log10 cfu/ml) levels in the milk samples were detected as 6.36, 2.95, 5.74, 1.10, 2.46 and 2.63, respectively.

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Introduction:-

Milk is a white liquid produced by the mammary glands of mammals. It is the primary source of nutrition in the world with its high nutritious property. If convenient storage temperatures are not paid attention, milk becomes a suitable propagation medium for microorganisms due to its biochemical composition and high water activity. Milk can be easily contaminated and spoiled when it is produced in unhygienic environment. Milk quality is directly related to its composition and hygiene (Oliver et al., 2005; Parekh and Subhash, 2008). Buffalo milk has become a research subject and received increasing attention in many countries due to its rich nutrient content (Amarjit and Toshihiko, 2003). Compared to cow milk, buffalo milk has a richer taste due to its contents of milk fat, protein, lactose, total dry matter, vitamin and minerals. These properties allow a wider variety for buffalo milk as raw material for milk products like cheese, butter and ice-cream (Fundaro et al., 2001). There has been an increasing demand for cheese made of buffalo milk in many countries throughout the world as it is an organic product (Bilal et al., 2006).

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CLASSIFICATION OF INDIAN BUFFALOSBuffaloes are grouped into riverin and swap types. Buffaloes in Turkey originate from Mediterranean buffaloes, a sub-group of river buffaloes and they are named BUFFALOes (Atasever and Erdem, 2008). it is reported that global buffalo milk was 90 million tons in 2009 and accounted for 13% of total milk production. The same report stated that more than 90% of total buffalo milk in the world is produced by India and Pakistan. (Spanghero and Susmel, 1996). Interest and investments in buffalo milk in different countries is increasing each year due to its unique taste and nutritious content (Amarjit and Toshihiko, 2003). It is reported that especially the products made of buffalo milk like mozzarella cheese, cream, ice-cream and yogurt have commercial impor-tance, as well (Fundaro et al., 2001). As in raw milk, microorganisms could rapidly propagate in buffalo milk due to rich nutrient content. In previous studies carried out on microbiological properties of buffalo milk (Sethi et al., 1994; Sekerden et al., 1999; Han et al., 2007), coliform bacteria, Escherichia coli, lactic acid bacteria, yeast-mold and Staphylococcus spp., were isolated from buffalo milk. Saprophytic microorganisms in milk could spoil buffalo milk, while the presence of pathogen bac-teria could pose a potential health threat

(Boycheva et al., 2002; Han et al., 2007). Contamination of milk and milk products mostly results from human factor and unhygienic conditions. Milk is generally contaminated in milk collection places. Coliform bacteria are microorganisms in natural flora of human and animal intestinal tract and they are accepted as indicators for bacteriological quality of milk and milk products (Chatterjee et al., 2006). Fur-thermore, the presence of these microorganisms indica-tes the possible existence of enteric pathogens that could threat public health. The most important indicators for microbiological quality include total bacteria number, coliform, yeast and mold quantity and detection of spe-cific pathogens and their toxins (Szita et al., 2008). Among all microorganisms, E. coli is an organism that frequently contaminates foods. They generally exist in milk and milk products due to inadequate sanitation (Jayarao and Henning, 2001). The main thing in micro-biological investigation of milk is the determination degree and number of indicator micro-organisms. Coliform bacteria are reported to define the suitability of milk for human consumption (Wells et al., 1991).

Besides microbiological quality of milk, its physical and chemical properties are also quite important. Changes in milk composition depend on many factors like genetic, lactation time, daily variations, birth, alimentation type, age, udder cleaning and season (Kilic and Kilic, 1994; Haenlein, 2003). These factors greatly affect the quality and processing ability of cheese, butter and other milk products (Lindmark-Manson et al., 2000; Barron et al., 2001; Lindmark-Manson et al., 2003). Geographic region, climate conditions and lactation period are known as seasonal changes and cited among factors affecting milk composition (Sethi et al., 1994; Suman et al., 1998; Sekerden et al., 1999; Boycheva et al., 2002; Waldner et al., 2002). Especially, there is an inverse proportion bet-ween ambient temperature and protein and fat content of milk. Solid fats tend to decrease with increasing air tem-perature. Fat, protein, casein and all fractions of nitrogen are affected by seasonal change (Ng-Kwai-Hang et al., 1984; Lacroix et al., 1996). It is reported that high am-bient temperature causes clot hardness to decrease and

Gürler et al. 1513 clot formation rate and clotting time to increase (Sevi et al., 2001). At high ambient temperatures, fat concentra-tion decreases, while lactation period extends (Kilic and Kilic, 1994; Sekerden, 1999). Some studies reported that significant changes occur in the amounts of short chain fatty acids, and they become minimum in winter and maximum in summer (Lindmark-Manson et al., 2003; Lock and Garnsworthy, 2003).

The main physical characteristics of milk are defined as pH and electrical conductivity; in addition, fat content of buffalo milk is the most variable milk component, which is caused by genetic and specific factors (Harmon, 1994; Hartet et al., 1999; Ma et al., 2000). Carbohydrates are preliminary form of glucose circulating as lactose in milk. Lactose level is reported to affect milk amount to be pro-duced and the level of short chain fatty acids (Fernando et al., 1983; Hamman and Gyodi, 1994). High fat and calorie contents besides dry matter are regarded as the superior and distinctive property of buffalo milk (Soysal, 2006). Buffalo milk is processed into many products including butter, cream, hard and soft cheese, ice-cream and yogurt (Bilal et al., 2006).

The aim of this study carried out in Afyonkarahisar Pro-vince, located in an important region for buffalo milk and milk products with dense buffalo presence, was to deter-mine the chemical and microbiological qualities of seasonally collected buffalo milk samples and to investigate the effect of seasonal change on milk composition.

Materials And Methods:-

Milk samples

A total of 120 raw milk samples were collected every month from 10 small sized family enterprises (farms had ≤ 10 buffaloes) randomly selected in beed 10 samples from each, between September 2012 and August 2013. Two hundred and fifty milliliter (250 ml) of milk samples were taken from producer under aseptic conditions and transferred in sterile bottles to laboratory in cold chain 4°C' and then they were analyzed.

Chemical analyses

Fat, protein, lactose, total dry matter (DM) and ash contents of buffalo milk samples were determined by precalibrated LactoStar milk analysis device (FUNKE GERBER, Germany). pH values of milk samples were determined by InoLab (pH Level L 01280054) pH meter device.

Microbiological analyses

In the analysis, 10 ml of milk was taken from each milk sample and homogenized in sterile bags containing 90 ml of sterile buffered peptone water for 1-2 min and then dilutions were prepared by 108 and cultivations were performed. Cultivation was made on Plate Count Agar (Oxoid CM0325) using drop plate method in order to determine the

number of mean total bacteria count (TCA) in the prepared dilutions (Anonymous, 2003). Violet Red Bile Agar (Oxoid-CM 107) was used to determine coliform bacteria count and incubated under aerobic conditions at 37°C for 24 h (Anonymous, 1993). Cultivation was made on MRS Agar (Oxoid-CM 0361) for lactic acid bacteria (LAB) count and left for 48-72 h of incubation under anaerobic conditions at 30°C (Gas Generation Kit Oxoid BR 0038) (Harrigan and MacCance, 1976).

Season	Total solids (%w/w)	Non fat solid	Fat (%v/v)	Protein (%w/w)	Lactose (%w/w)	Ash (%w/w)	pН
		(%w/w)			, , ,	`	
Winter	17.41±0.54a	8.89±0.28a	7.67±0.55a	4.90±0.26a	3.99±0.12b	0.78±0.07a	6.68±0.11a
(n=30) Spring	15.61±0.74c	8.49+0.23b	6.38+0.66b	4.03+0.16c	4.45+0.20a	0.69+0.07b	6.45±0.06c
(n=30)	10.01_0.710	0.1920.200	0.0020.0000	1100_01100	1110_01200	0.09_0.070	0.15_0.000
Summer (n=30)	15.90±1.03c	8.49±0.27b	6.70±0.89b	4.05±0.14c	4.43±0.25a	0.66±0.06b	6.45±0.12c
Autumn (n=30)	16.59±0.57b	8.37±0.25b	7.40±0.51a	4.46±0.18b	3.91±0.21b	0.77±0.07a	6.55±0.12b
Mean (120)	16.38 ± 1.01	8.56±0.32	7.04±0.84	4.36±0.40	4.19±0.32	0.72±0.08	6.55±0.13

Table 1:- Chemical analysis results of BUFFALO milk.

Cultivations were made on Tryptone Bile X- Glucuronide Medium (TBX) (Oxoid-CM945) for E. coli count and incubated at 44°C for 18-24 h (Anonymous, 2001). Egg Yolk Tellurite emulsion (Oxoid-SR54) was included for S. aureus count and performed in Baird Parker Agar (Oxoid -CM275) at 37°C for 24-48 h aerobic conditions (Anonymous, 1999). Cultivations were made on Rose Bengal Chlo-ramphenicol Agar (Oxoid-CM549) for yeast and mold count and incubated at 25°C for 72 h under aerobic conditions (Anonymous, 1987).

Statistical analyses

The microorganism numbers detected in this study were transferred to base 10 logarithm values and then statistical data was obtained by SPSS statistics software. One way variance analysis was per-formed to determine the differences between microorganism num-bers and chemical parameters in terms of seasons, and Duncan test was applied to determine differences among the means.

Results:-

Chemical analysis

As a result of the chemical analyses of milk samples col-lected from buffaloes in different seasons, the total solids (TS) (16.38% \pm 1.01), non-fat solids (NFS) (8.56 \pm 0.32), fat (7.04% \pm 0.84), protein (4.36% \pm 0.40), lactose (4.19% \pm 032), ash (0.72% \pm 0.08) and pH (6.55 \pm 013) values were determined (Table 1). Generally milk yield was determined to increase in plains, humid and rainy areas, while fat content was found higher in cold and mountainous areas. The data of this study indicated that milk fat and protein contents increase concurrently, while TS, NFS, fat, protein, ash and pH values are highest in winter, while TS, fat and protein levels are lowest in spring.

Microbiological analysis

Regardless of seasons, the mean total bacteria count (TBC), coliform bacteria, lactic acid bacteria (LAB), E. coli, S. aureus and yeast-mold values (log10 cfu/ml) were determined as 6.36 ± 0.28 , 2.95 ± 0.21 , 5.74 ± 0.31 , 1.10 ± 0.17 , 2.46 ± 024 and 2.63 ± 0.25 , respectively (Table 2).

Contamination levels of milk with TBC, coliform, S. aureus and yeast-mold were found lower in winter than in other seasons, while the lowest E. coli contamination was detected in autumn. The highest contamination levels with these bacteria were observed in summer. Microbio-logical values of milk samples were found at similar levels in spring and autumn.

Discussion:-

In this study, the mean total solids, non-fat solids, fat, protein, lactose and pH levels of milk samples collected in different seasons were determined as 16.38%, 8.56%, 7.04%, 4.36%, 4.19%, 0.72% and 6.55, respectively. Fat, protein and ash contents were determined to decrease in hot summer months, while lactose content was reported to

increase (Yöney, 1974). It was reported that buffalo milk contain has higher nutritious values with higher protein, fat, lactose and TS than cow milk (Fundora et al., 2001; Lidmark-Manson et al., 2003). Different resear-chers reported that alimentation, lactation period, milking frequency, milking method and season have important effects on physicochemical parameters of buffalo milk (Sethi et al., 1994; Suman et al., 1998; Sekerden et al., 1999; Boycheva et al., 2002; Waldner et al., 2002).

In this study, the mean lactose level of buffalo milks $(4.19\% \pm 032)$ was found lower than those reported by Han et al. (2007) and Najdenova and Dimitrov (2003). On the other hand, milk fat is the most changeable milk component. Fat content is affected by many factors. The most important factors are seasonal change and lactation period. Fat, protein and ash contents tend to increase in winter, and milk yield is reported to increase in the later periods of lactation, while fat and protein contents de-crease. It is also reported that habitat and feeding pattern are quite effective on milk fat and protein levels, and milk protein and NFS contents of animals grazing in summer are higher than those of animals closed-fed in winter (Yöney, 1974; Sevi et al., 2004). In this study, the mean fat (7.04 \pm 0.84 %v/v), total solids (16.38 \pm 1.01% w/w) and protein (4.36 \pm 0.40% w/w) contents are slightly lower than those reported by Najdenova and Dimitrov (2003) and Han et al. (2007). Similar results were reported by Sarfarz et al. (2008) and Çelik et al. (2001), but the values of the present study were higher than found by Kanwal et al. (2004). On the other hand, Ariota et al. (2007) reported fat and protein contents of buffalo milk as 8.71 and 3.86%, respectively, and pH as 6.58.

The mean ash values $(0.72 \pm 0.08\% \text{ w/w})$ of BUFFALO milk are similar to the values reported by Celik et al. (2001) and Sarfarz et al. (2008), but higher than the value found by Sekerden and Avsar (2008). The mean pH (6.55 ± 0.13) of BUFFALO milk is similar to the values reported by Han et al. (2007) and Sekerden and Avsar (2008), but higher than those of Aurelia et al. (2009).

Variance analysis was used to investigate chemical composition. Total solids, protein, non- fat solids and pH values were significantly higher (p<0.01) in winter than in other seasons. On the other hand, the lactose content was found higher (p<0.01) in spring and summer than in winter and autumn. The mean fat and ash contents were found highest in winter and autumn.

Similar to milk components, microbiological quality of milk changes by ambient temperature. In this study, the mean total bacteria count (TBC), coliforms, lactic acid bacteria, E. coli, S. aureus and yeast-mold (log10 cfu/ml) levels were determined as 6.36 ± 0.28 , 2.95 ± 0.21 , 5.74 ± 0.31 , 1.10 ± 0.17 , 2.46 ± 024 and 2.63 ± 0.25 , respec-tively, and microorganism load was determined to increase in warm months.

The Turkish National Standard (TSE 1018) for TBC was $1.0 \ge 100$ x 105 cfu/ml for raw cow milk. The EU specifica-tion (EU Directive 92/46/EEC, 2004) for raw buffalo milk is an average of $5 \ge 105$ cfu/ml TC. In this study, the mean Gürler et al. 1515 TBC was determined as $2.30 \ge 100$ cfu/ml. This level is higher than both the TSE and EU standards. The main reason for these relatively higher counts of TBC should be ascribed to poor hygiene conditions during milking, collection and transport.

S. aureus may access bulk milk either by direct excre-tion from the udder with clinical and subclinical staphylococcal mastitis, or by fecal contamination (Callon et al., 2008). Interchange of staphylococcal strains and poor microbiological quality of raw milk may be attributed to skin particles in the environment and poor sanitary practice (Normanno et al., 2007).

In another study carried out in China, TBC, LAB, yeast-mold, coliforms, E. coli and S. aureus (log 10 cfu/ml) levels in 120 buffalo milk samples were determined as 5.59 4.62, 1.79, 2.42, 1.53 and 1.68, respectively (Han et al., 2007). Coroian et al. (2010) reported mean coliform bacteria, yeast-mold and aerobe mesophile general crea-ture levels in 42 Romanian buffalo milk samples as $4.96 \pm 0.45/\text{ml}$, $633.47 \pm 0.01/\text{g}$ and $4.46 \pm 0.11 \times 105/\text{ml}$, respectively and they also determined 3.27 log cfu/ml of E. coli in three samples. According to the results of this study, coliform, E. coli and yeast-mold levels were lower than those reported by Coroian et al. (2010); however, TBC level was found higher. The same researchers carried out a study on Murrah buffalo species and deter-mined coliform, E. coli, S. aureus and yeast-mold levels (log10 cfu/ml) as 3.95 ± 0.07 , 1.80 ± 0.23 , 1.80 ± 0.23 and 1.33 ± 0.46 , respectively. Accordingly, coliform and E. coli levels were lower than those determined in the present study, while S. aureus and yeast-mold levels were hig-her. Similarly, Desmasures et al. (1997) studied cow milk in different seasons, and reported that TBC, LAB, yeast, coliform and S. aureus levels (log10 kob/ml) were 7.1x103, 1.8x102, 7.2x101, 5.7x101 and 4.5x102, respectively, in winter, while these levels were determined as 8.6x103, 1.8x102, 8.4x101, 7.7x101 and 3.5x102 in summer; accor-dingly, the levels determined in the present study in win-ter period were lower than those reported by Desmasures et al. (1997). Considering summer levels, TBC was found high, while others were low. Similarly, Ali et al. (2010) de-termined mean TBC, LAB, coliform, E. coli and S. aureus levels in cow milk as 5.86 ± 0.31 , 4.47 ± 0.44 , 2.76 ± 0.18 , 1.63 ± 0.20 and 1.92 ± 0.47 , respectively.

In another study on raw buffalo milk samples, TBC, E. coli, and yeast levels (log10 cfu/ml) were determined between 3.4 x 105-4.0 x 107, 2.0x101-1.7 x 104 and 2.7 x 102-1.7 x 104, respectively (Braun and Preuss, 2007) compared with this study. LAB constituted a major part of the microflora with an average of 5.74 ± 0.31 log cfu/ml (Han et al., 2007) and mean LAB level in buffalo milk was reported as 4.62 ± 0.12 log cfu/ml (Lingathurai et al., 2009) and 4.46 ± 0.44 cfu/ml, while it was found 4.47 ± 0.44 cfu/ml in raw cow milk (Ali et al., 2010). Boycheva et al. (2002) observed that LAB was predominant in Bulga-rian buffalo milk. High level of LAB in raw milk would result in undesired fermentative acidity, and it is sugges- 1516 Afr. J. Microbiol. Rested to take effective precautions for preventing this kind of spoilage (Han et al., 2007).

Chemical composition of buffalo milk provides perfect opportunities for the development of local milk industry and providing nutrient element needs of humans. In addi-tion, the presence of pathogens, indicators and index microorganisms in raw milk and products made of inade-quately heat-treated milk could pose a threat for public health.

Livestock enterprises making milk production are com-posed of family enterprises in villages and towns with large numbers. Small sized production units have diffi-culty in obtaining inputs and services like adequate shel-ter, feed, technical information, veterinary services for buffalo and cow dairy production. It is possible to preci-sely organize hygienic and technological stages from production to consumption of milk and milk products only when all potentials are combined. In this regard, the prin-cipal that quality milk comes from healthy udder, healthy animal and clean environment gains great importance.

The important rule in food processing is the good quality of raw material. A good quality of end product can-not be obtained from a raw material with poor hygienic quality. Spoilage process of milk starts with milking. The previous studies confirmed that milk hygiene does not receive enough attention in Turkey. Considering the unhealthy conditions in milk production and other conta-mination sources in milk processes, milk can be a con-veyor of pathogens threatening public health. Microorga-nisms cause rapid souring, spoilage and undesired color, taste and bitterness in milk and thus resulting in poor quality. It is reported that many epidemic disorders of milk origin are caused by dirty hands of workers in milk pro-duction, dirty tools and equipment, insects and dirty water sources. Provision of microbiological quality parameters of raw milk and milk products plays an important role in quality control. It is necessary to minimize technological and economic losses in milk processing and obtain a longer shelf life.

References:-

- 1. Ali AA, Irshad N, Razaz SA, Manahil AA (2010). Microbiological safety of raw milk in Khartoum state, Sudan: 1-Khartoum and Omdurman cities. Pak. J. Nutri., 9(5): 426-429.
- 2. Amarjit SN, Toshihiko N. (2003). Role of buffalo in the socioeconomic development of rural Asia: Current status and future prospectus. Anim. Sci. J., (74): 443–445.
- 3. Ariota B, Campanile G, Potena A, Napolano R, Gasparrini B, Neglia GL, DiPalo R (2007). Ca and P in buffalo milk: curd yield and milk clotting parametters. Ital. Anim. Sci., 6(6): 497-499.
- 4. Bilal MQ, Suleman M, Raziq A (2006). Buffalo: Black Gold of Pakistan. Livestock Res. Rural Development, 18(9): 128.
- 5. Chatterjee SN, Bhattacharjee I, Chatterjee SK, Chandra G (2006). Microbiological examination of milk in Tarakeswar, India with special reference to coliforms. Afr. J. Biotechnol., 5: 1383-1385.
- 6. Coroian A, Coroion CO, Vodnar DC, Trif M (2010). Study on the main microbiological traits in Romanian buffalo milk. Bioflux, 2(2): 92-98.
- 7. Fernando RS, Spahr SL, Jaster EH (1983). Comparacion of electrical conductivity of milk with other indirecty methods for detection of subclinical mastitis. J. Dairy Sci., 68: 449-456.
- 8. Kanwal R, Ahmed T, Mirza B (2004). Comparative analsis of quality of milk collected from Buffalo, Cow, Goat and Sheep of Rawalpindi/Islamabad region in Pakistan. Asian J. Plant Sci., 3(3): 300-305.
- 9. Kilic A, Kilic S (1994). Feeding and milk. Bilgehan Press, Izmir, 1994.

- Najdenova N, Dimitrov T (2003). Technological qualities of buffalo milk from the Bulgarian Murrah breed for production of Bulgarian yoghurt. J. Anim. Sci., 40(5): 33–35.
- 11. Ng-Kwai-Hang, KF, Hayes JF, Moxley JE, Monardes HG (1984). Variability of test-day milk production and composition and relation of somatic cell counts with yield and compositional changes of bovine milk. J. Dairy. Sci., 67: 361-366.
- 12. Parekh TS, Subhash R (2008). Molecular and bacteriological examination of milk from different milch animals with special reference to coliforms. Current Research in Bacteriology, 1: 56-63
- 13. Sarfarz A, Gaucher I, Rousseau F, Beaucher E, Piot M, Grongnet JF, Gaucheron F (2008). Effect of acidification on physico-chemical charecteristics of buffalo milk: A comparison with cow's milk. Food Chem., 106 (1): 11-17.
- 14. Waldner DN, Stokes SR, Jordan ER, Looper ML (2002). Managing milk composition: Normal sources of variation.
- 15. Wells JG, Shipman LD, Greene KD, Sowers EG, Green JH, Cameron DN, Downes FP, Martin ML, Griffin PM, Ostroff SM (1991). Isolation of Escherichia coli serotype O157:H7 and other Shiga-like-toxin-producing E. coli from dairy cattle. J. Clin. Microbiol., (29): 985–989.
- 16. Yöney Z (1974). Milk Chemistry. Ankara Univ. Agric. Fac. Publication, No. 530, Textbook: 135, Ankara.