



RESEARCH ARTICLE

Hematotoxicity induced by sub-chronic dose of 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin in Swiss albino mice (*Mus musculus*)*Tanuja^{1,2}, Ruchi², Anjali Singh³, and J.K. Singh⁴

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Abstract

Present study was designed to determine the acute oral toxicity (LD₅₀) and investigate the effect of sub-chronic dose (0.5µg/Kg body wt.) of 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin (TCDD) on the haematological parameters in Swiss albino mice. The LD₅₀ was found 5.087 µg/Kg body wt. in Swiss albino mice. The effect of sub-chronic dose of TCDD on the haematological parameters showed statistically very significant ($P<0.01$) decrease in the levels of RBC (red blood cell), Hb (Haemoglobin), MCV (Mean Corpuscular Volume), MCH (Mean Corpuscular Haemoglobin), MCHC (Mean Corpuscular Haemoglobin Concentration), and HCT (Hematocrit). Whereas, statistically significant ($P<0.05$) increase in the level of WBC (White blood cell) was observed as compared to control group. The research finding suggests that exposure of sub-chronic dose of TCDD significantly altered the haematological parameters, which is relevant to risk evaluation of TCDD that may be used as a predictive values for its toxicity in mammalian system including human being.

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INTRODUCTION

The presence of toxic pollutant in environment is the result of increased anthropogenic activity. Among the pollutant released, 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin (TCDD) is a widespread, persistent, and highly toxic. It is formed as an unintentional by-product of incomplete combustion and various chemical processes (Ziegler, 1997). Major sources of environmental dioxin pollution are herbicide and fungicide manufacturing, the paper and cellulose industry, thermal reactions of chlorinated aromatic compounds, and transformer or condenser break-down (Kulkarni et al., 2008). It is persistent, slowly degradable in environment, and generates toxicity for a longer duration of time (ten-Tusscher et al., 2000). It causes adverse effect like haematological disturbances, malignancy, congenital malformation, and immune suppression etc. (Vos et al., 1997; Neubert et al., 1993) at different doses and at different period of time. It has also been reported that acute and sub-acute exposure of TCDD on mammal causes undesirable effects like hepatotoxicity, teratogenicity, interference with lipid metabolism, chloracne, neurobehavioral disturbance, endocrine disruption, wasting syndrome, and reproductive toxicity (Schechter et al., 2006; Senft et al., 2002; Pluim et al., 1994).

Analysis of blood parameters is relevant to risk evaluation as the predictive value for toxicity. Haematological parameters analysis have been widely used in the diagnosis of a range of diseases induced by industrial compounds,

drugs, dyes, heavy metals, pesticides, and several others (Badraoui et al., 2011; Olson et al., 2000), as blood being a medium for the transport of intercellular and intracellular materials.

Therefore, in the present study, an effort has been made to determine acute oral toxicity (LD_{50}), and sub-chronic exposure of TCDD on some of the haematological parameters in Swiss albino mice (*Mus musculus*) such as red blood cell (RBC) and white blood cell (WBC), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), haemoglobin (Hb), and hematocrit (HCT) were monitored.

Material and Methods

Test Chemicals

The test chemical, TCDD (2,3,7,8-Tetrachlorodibenzo-*p*-dioxin) was purchased from Accustandard, USA (CAS 1746-01-6; molecular weight, 321.9; purity, 99% as analyzed by GC-MS). It was dissolved in corn oil (Sigma Chemical Co., St. Louis and Neishel chemical Pvt. Ltd., India) which was used as vehicle.

Experimental animal

Healthy Swiss albino mice of both sexes, weighing between 30-35g were obtained from a random-bred colony at the Mahavir Cancer Sansthan, Patna, Bihar, India. They were maintained under the optimal condition at the temperature ($24 \pm 1^\circ\text{C}$), humidity ($55 \pm 5\%$), and lighting (12-h light/dark cycle). Food and tap water were given *ad libitum* during the study. Study were carried out as per CPCSEA guidelines (Approval No.-1129/bc/07/CPCSEA).

Acute toxicity determination (LD_{50})

Six experimental groups of mice have been formed with six mice in each group. The groups were numbered as Group-1 to Group-6. All the groups were treated at different concentration of TCDD for seven consecutive days. The distribution of doses to each treated groups has been summarized as: Group-I ($10\mu\text{g/Kg}$ body wt.), Group-II ($8\mu\text{g/Kg}$ body wt.), Group-III ($6\mu\text{g/Kg}$ body wt.), Group-IV ($3\mu\text{g/Kg}$ body wt.), Group-V ($2\mu\text{g/Kg}$ body wt.), and Group-VI ($1\mu\text{g/Kg}$ body wt.).

Sub-chronic toxicity study

Based on the finding of LD_{50} , sub-chronic dose was selected as $0.5\mu\text{g/Kg}$ body wt. and administered to the mice for 2 and 4 weeks. Treatment volume was determined on the basis of body weight. After acclimatization for a week experimental mice were again divided into three groups and classified as: Group-I (Normal) received food and tap water *ad libitum*, Group-II (Control) received corn oil in addition to food and tap water *ad libitum*, and Group-III (Treated) received sub-chronic ($0.5\mu\text{g/Kg}$ body wt.) dose of TCDD dissolved in corn oil (vehicle) in addition to food and tap water *ad libitum*. Each group consists of six mice and repetition was made three times.

Haematological Parameters

Blood samples were collected from normal, control and sub-chronic dose of TCDD administered groups of mice at intervals of 2 and 4 weeks by ocular vein puncture for the determination of haematological parameters. The estimation of the levels of red blood cell (RBC), white blood cell (WBC), haemoglobin (Hb), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), and hematocrit (HCT) were done by using EDTA (ethylenediaminetetraacetate) as an anticoagulant. Haematological parameters were assessed using Cell Counter (Medonic M-16 Series) in the Department of Haematology, Mahavir Cancer Sansthan, Patna (Bihar).

Statistical analysis: Data was analyzed and experimental values were expressed as the mean \pm SEM and *P* value was calculated using one way analysis of variance (ANOVA) by using SPSS (11.5 version) software. $P < 0.05$ was considered statistically significant and $P < 0.01$ was considered statistically very significant.

Result and Discussion

In the present study the acute oral toxicity (LD_{50}) of the TCDD was found to be $5.087\mu\text{g/Kg}$ body wt. using Probit analysis (EPA PROBIT ANALYSIS PROGRAM, used for calculating LC/EC value, version 1.5) (Table-1). However, acute oral toxicity study in rat was found to be $20\mu\text{g/Kg}$ body wt. (NTP, 2000).

The present study indicates the hematotoxic effects of sub-chronic dose of TCDD in Swiss albino mice. The decrease in the levels of RBC, Hb, MCV, MCH, MCHC and HCT were observed and found to be statistically very ($P<0.01$) significant (Table-2). Similar kind of reduction in the haematological parameters due to TCDD has also been reported in a study on human, guinea pigs, female rats, Zebrafish, goats, and mice (Eraslan et al., 2009; Uboh et al., 2012; Li et al., 2010; Belair et al., 2001; Carney et al., 2006; Leijs et al., 2008; Chu et al., 2007). Decrease in the levels of RBC, Hb, and HCT have also been observed in the case of other persistent organic pollutant due to TBDD dioxin in both sexes of rats (Seigo et al., 2006).

Decrease in the levels of RBC and Hb may be due to the altered haematopoiesis (Badraoui et al., 2011), that results in excessive destruction or decrease in the synthesis of RBC. As a result of destruction of RBC, free radicals are generated (Fibach et al., 2008; Stohs, 1990; Bassem et al., 2008) and caused oxidative stress. Free radicals induced the production of reactive oxygen species, reactive nitrogen species, and lipid per-oxidation (Zahran, 1997). This ultimately results into RBC cell membrane damage, and caused anemia (Helliwell and Mediline, 1994). Disorders of erythropoiesis and Hb synthesis may be the consequence of the endocrine disorders (Geusau et al., 2001).

A very significant ($P<0.01$) decrease in levels of MCV, MCH, and MCHC were observed in this study. The decrease in the levels of MCH, MCHC and MCV may be due to the failure in blood osmoregulation and plasma osmolarity (Pohjanvirta et al., 1989). The levels of MCH, MCHC, and MCV found in this study represent the characteristic features of anaemic condition in the treated groups.

However, observation shows that the level of WBC increased significantly ($P<0.05$) in the sub-chronic dose of TCDD administered groups (Group-III) as compare to control Group-II. It may be due to an increase in phagocytic activity towards destructive RBCs, resulting into activation of immune system of the body by innate defence capability of mice against toxins (TCDD). Reduction in erythrocytes and increase in WBC possibly directly associated with marked increase in aryl hydrocarbon receptor (AhR) expression to mediate toxic effect (Crawford et al., 1997). Haematological alterations have also been observed due to Dioxin (TCDD) in human with the decreased level of erythrocytes and increased level of WBC (Geusau et al., 2001).

From the present investigation it can be inferred that at low dose and at less time duration, TCDD is capable of inducing significant alteration in haematological parameters of the experimental animal model Swiss albino mice.

Table-1: Environmental Protection Agency PROBIT analysis program used for calculating LC/EC values (Version 1.5) for TCDD			
Concentration of TCDD	No of subject	Observed response	Expected response
10 µg/Kg body wt.	6	5	4.722
8 µg/Kg body wt.	6	4	4.219
6 µg/Kg body wt.	6	3	3.463
3 µg/Kg body wt.	6	2	1.601
2 µg/Kg body wt.	6	1	0.814
1 µg/Kg body wt.	6	0	0.166
Estimated LC/EC 50.0 of TCDD was 5.087 µg/Kg body wt.			

Table-2: Effect of oral administration of sub- chronic dose of TCDD on haematological profile of Swiss albino mice						
Sl.No.	Haematological parameters	Group-I	2 Weeks		4 Weeks	
			Group-II	Group-III	Group-II	Group-III
1.	RBC($10^6/\text{mm}^3$)	9.106±0.078	9.32±0.116	8.306**±0.041	9.20±0.11	7.24**±0.21
2.	MCV(μm^3)	41.06±0.118	41.52±0.216	37.90**±0.087	41.335±0.1321	35.9**±0.104

3.	WBC($10^3/\text{mm}^3$)	5.50±0.097	5.329±0.023	9.385*±0.090	5.267±0.021	13.2*±0.042
4.	Hb(g/dl)	14.43±0.176	14.54±0.23	9.383**±0.081	14.9±0.012	8.89**±0.042
5.	MCH(Pg)	15.23±0.012	15.02±0.094	14.18**±0.056	15.12±0.024	11.2**±0.031
6.	MCHC(g/dl)	36.2±0.023	34.0*±0.035	30.62**±0.065	33.46*±0.036	31.43**±0.11
7.	HCT(%)	38.42±0.012	38.64±0.175	31.45**±0.116	37.1*±0.85	29.6**±0.42

Route of administration: oral; values expressed as mean±SEM (n=6); *Significant ($P<0.05$): **very significant ($P<0.01$) as compared to control (Group-II); RBC: Red blood cell; MCV: Mean corpuscular volume; WBC: White blood cell; Hb: Hemoglobin; MCH: Mean corpuscular hemoglobin; MCHC: Mean corpuscular hemoglobin concentration; HCT: Hematocrit; Group-I: normal; Group-II: control (corn oil treated); Group-III: treated (sub-chronic dose of TCDD).

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References

- Badraoui, R., Abdelmoula, B. and Rebai, T. (2011).** Erythrocytes oxidative damage and haematological effects of 2,4,4,5-tetrachlorodiphenyl sulfone in rats. *Exp Toxicol Pathol.* 63: 479-485.
- Bassem, M.R. and Fouzy, A.S.M. (2008).** Oxidative Stress in Rats Exposed to 2,3,7,8-Tetrachlorodibenzo-*p*-Dioxin with Emphasis on the Magnetic Properties of Iron Atom in Hemoglobin. *Global Veterinaria.* 2: 250-256.
- Belair, C.D., Peterson, R.E. and Heideman, W. (2001).** Disruption of erythropoiesis by dioxin in the zebrafish. *Dev Dyn.* 222: 581-94.
- Carney, S.A., Prasch, A.L., Heideman, W. and Peterson, R.E. (2006).** Understanding dioxin developmental toxicity using the zebrafish model. *Birth Defects Res A Clin Mol Teratol.* 76: 7-18.
- Chu, I., Valli, V.E. and Rousseaux, C.G. (2007).** Combined effects of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin and polychlorinated biphenyl congeners in rats. *Toxicol Environ Chem.* 89: 71-87.
- Crawford, R.B., Holsapple, M.P. and KAMINSKI, N.E. (1997).** Leukocyte activation induces aryl hydrocarbon receptor upregulation, DNA binding, and increased cyp1a1 expression in the absence of exogenous ligand. *Mol Pharma.* 52: 921-927.
- Eraslan, G., Kanbur, M., Silici, S., Liman, C.B., Altinordulu, S. and Sarica, Z.S. (2009).** Evaluation of protective effect of bee pollen against propoxur toxicity in rat. *Ecotoxicol Environ Saf.* 72: 931-937.
- Fibach, E. and Rachmilewitz, E. (2008).** The role of oxidative stress in haemolytic anaemia. *Curr Mol Med.* 8: 609-619.
- Geusau, A., Abraham, K., Geissler, K., Sator, M.O., Stingl, G. and Tschachler, E. (2001).** Severe 2,3,7,8-Tetrachlorodibenzo-*p*-dioxin (TCDD) Intoxication: Clinical and Laboratory Effects. *Environ Health Perspect.* 109: 865-869.

- Helliwell, B. and Mediline. (1994).** Free radicals and antioxidants: a personal view. *Nutr Review.* 52: 253-265.
- Kulkarni, P.S., Crespo, J.G. and Afonso, C.A.M. (2008).** Dioxins sources and current remediation technologies – a review. *Environ Int.* 34: 139-153.
- Leijs, M.M., ten-Tusscher, G.W., Olie, K., Aalderen, W.M.C., Vulsma, T., Westra, M., Oosting, J. and Koppe, J.G. (2008).** Perinatal dioxin exposure in the Netherlands; a long term follow up. *Int J Environ Heal R.* 2: 429-438.
- Li, W., Vogel, C.F., Wu, D. and Matsumura, F. (2010).** Non-genomic action of TCDD to induce inflammatory responses in HepG2 human hepatoma cells and in liver of C57BL/6J mice. *Biol Chem.* 391: 1205-1219.
- Neubert, R., Stahlmann, R., Korte, M., Van, L.H., Vos, J.G., Golor, G., Webb, J.R., Helge, H. and Neubert, D. (1993).** Effects of small doses of dioxins on the immune system of marmosets and rats. *Ann N Y Acad Sci.* 685: 662-686.
- Olson, H., Betton, G., Robinson, D., Thomas, K., Monro, A., Kolaja, G., Lilly, P., Sanders, J., Sipes, G., Bracken, W., Dorato, M., Van-Deun, K., Smith, P., Berger, B. and Heller, A. (2000).** Concordance of the Toxicity of Pharmaceuticals in Humans and in Animals. *Regul Toxicol Pharm.* 32: 56-67.
- Pluim, H.J., Koppe, J.G., Olie, K., Slikke, J.W., Slot, P.C., Van, B. and Van, C.J. (1994).** Clinical laboratory manifestations of exposure to background levels of dioxins in the perinatal period. *Acta Paediatr.* 83: 583-587.
- Pohjanvirta, R., Kulju, T., Morselt, A.F., Tuominen, R., Juvonen, R. and Rozman, K. (1989).** Target tissue morphology and serum biochemistry following 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) exposure in a TCDD susceptible and a TCDD-resistant rat strain. *Fundam Appl Toxicol.* 12: 698-712.
- Schechter, A., Birnbaum, L., Ryan, J.J. and Constable, J.D. (2006).** Dioxins: an overview. *Environmental research.* 101: 419-428.
- Seigo, Y., Kasuke, N., Hideki, S., Tetsuya, T., Michiharu, M., Hisao, O.T.N., Kazunori, Y., Heihachiro, A. and Taijiro, M. (2006).** Systemic and myelotoxic effects of single administration of 2,3,7,8-tetrabromodibenzo-p-dioxin in rats. *Environ Health Prev Med.* 11: 136-44.
- Senft, A.P., Dalton, T.P., Nebert, D.W., Genter, M. B., Hutchinson, R.J. and Shertzer, H.G. (2002).** Dioxin increases reactive oxygen production in mouse liver mitochondria. *Toxicol. Appl. Pharmacol.* 178(1): 15-21.
- Stohs, S.J. (1990).** Oxidative stress induced by 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD). *Free Radical Bio Med.* 9: 79-90.
- ten-Tusscher, T.G.W., Stam, G.A. and Koppe, J.G. (2000).** Open chemical combustions resulting in a local increased incidence of orofacial clefts. *Chemosphere.* 40: 1263-1270.
- Uboh, F.E., Usuh, I.F., Nwankpa, P. and Obochi, G.O. (2012).** Effect of oral exposure to nitrocellulose thinner on haematological profiles of male albino wistar rats. *Am J Biochem Mol Biol.* 2: 227-234.
- NTP (2000).** U.S. National Toxicology Program acute toxicity studies for Dioxin (2,3,7,8-TCDD).
- Vos, J.G., De-Heer, C. and Loveren, V. (1997).** Immunotoxic effects of TCDD and toxic equivalency factors. *Teratog Carcinog Mutagen.* 17: 275-84.
- Zahran, W.M. (1997).** Sracoptic manage haematological alterations, skin lesions and hypersensitivity in naturally infested rabbits. *J.Union Arab Biol Cairo Zool.* 11: 136-144.
- Ziegler, J. (1997).** Environmental endocrine disruptors get a global look. *J Natll Cancer Inst.* 89: 1184-1187.