

Journal homepage: http://www.journalijar.com

INTERNATIONAL JOURNAL OF ADVANCED RESEARCH

RESEARCH ARTICLE

DROUGHT ALLEVIATION USING GLUTATHIONE IN CANOLA PLANTS

Sharbat L.M.*, Mehanna H.M., and M.S. Gaballah*

^{*}Water Relations & Field Irrigation Dept, Agric. Div., National Research Centre, Cairo, Egypt

.....

Abstract

.....

Manuscript History:

Manuscript Info

Received: 19 July 2014 Final Accepted: 26 August 2014 Published Online: September 2014

Key words:

Canola (Brassica nupas L.) -Irrigation intervals-Glutathione-Growth- Yield- Osmotic pressureproline-carbohydrate

*Corresponding Author

•••••

Sharbat L.M

..... Drought is one of most important environmental factors inhibiting photosynthesis and decreasing growth and productivity of plants. A Pot experiment was conducted in the greenhouse of the National Research Centre to evaluate the effect of glutathione GHS100 and 200 ppm, on Canola plants grown under different irrigation levels .Growth, yield, some chemical constituents and osmotic pressure (OP) were tested in plants. Plant height, number of green leaves, leaf area, and total dry weight positively responded to GHS treatments when canola plants received water every 7 days. Number of seeds and yield of seeds/plant responded to the interaction between irrigation intervals and glutathione spraving with 100 ppm glutathione was more effective than 200 ppm under 7 and 14 days irrigation intervals. The highest value of carbohydrate was obtained by spraying 100 ppm GHS and irrigation with 7 days intervals. Oil percentage only increased with application 100 and 200 ppm GHS under 7 days intervals. Application of glutathione showed slight increase in osmotic pressure of canola plants and 100 ppm GHS increased proline under 14 days irrigation intervals.

Copy Right, IJAR, 2014,. All rights reserved

Introduction

Brassica nupas (Canola) is a member of Brasicaceae which is covered with more bloom than other species in this family. Canola has been especially developed for oil by the Canadian scientists, where they reduced the amount of erucic acid in the newly bred variety. Canola constitutes low saturated fatty acids (6%) and high non-saturated fat. Canola has 50% less saturated fat than corn oil (Wiess, 1983). Canola cultivated area in Egypt is relatively very small due to the strong competition in cropping system with the strategic crops during winter season. It could be successfully cultivated in the newly reclaimed soils outside of Nile valley and delta areas for narrowing the gap of edible oil (Gallab and Sharaan, 2002 and Mekki, 2013). Drought strikes plants when transpiration rate is greater than rate of water absorption (Bray, 1997). Damages to plant caused by drought stress are variable depending on the level and duration of the stress and environmental factors (Glantz, 1994). Deficit water strategy is one of the management practices for coping with drought and shortage of water in arid and semiarid regions (Shabani, et al. 2013). To cope with water scarcity different approaches are proposed to decrease water consumption and increase water use efficiency, These methods included use of high yielding varieties (Mekki, 2013), fertilization (Bybordi and Ebrahimian (2013), growth regulators (Ullah, et al. 2012) or antioxidant application (Sakr and Arafa, 2009). Under both natural and agricultural conditions plants are often exposed to various environmental stresses. Drought is one of most important environmental factors inhibiting photosynthesis and decreasing growth and productivity of plants. It is one of the major causes of crop loss worldwide, reducing average yields for most major crop plants by more than 50% (Wang, et al. 2003).

Glutathione (GHS) is a tripeptide with a gamapeptide linkage between the amino group of cestine and the carboxylic group of glutamate side chain (Pompella, et al. 2003). As prototype antioxidant, has been involved in cell protection in noxious effect of excess oxidant stress both directly or cofactor of glutathione peroxidases. Glutathione is one of the major endogenous antioxidants in plants, known to play an important role in plant defense mechanisms. Glutathione functions as a substrate in antioxidative defense mechanisms by conjugating to toxic electrophilic

compounds, scavenging free radicals, and reducing peroxides. Several important catalytic enzymes that utilize glutathione in defense mechanisms, such as glutathione S-transferase (GST) and glutathione reductase (GR) show differential patterns of activity in plant tissues exposed to environmental stress conditions such as; cold, drought and wounding treatments (James and Davis, 2004 and Yılmaz, 2006).

Therefore, the objective of this work is to investigate the effect of glutathione on growth, yield, some chemical constituents and osmotic pressure in leaves of canola plants grown under different irrigation intervals.

MATERIALS AND METHODS

Pot experiment was conducted in the green net of the National Research Centre during the winter season of 2012-2013 to evaluate the effect of spraying glutathione on growth, yield and some chemical constituents and osmotic pressure in leaves of canola plants grown under different irrigation intervals. The treatments were as follows:

Irrigation intervals: Plants irrigated every week (7 days), 2 weeks (14 days) and three weeks (21 days). **Glutathione**: Spraying canola plants with 100 and 200 ppm and control plants sprayed with distilled water. The experiment included 9 treatments 3 irrigation intervals treatments and three glutathione treatments, with 8 replicates. Seeds of canola (Prassica nupus L) variety Serw 4 were sown on the 1st of Dec., 2012 and in earthenware pots (72 pots) filled with 12 kg of sandy loam soil. The physical and chemical properties of this soil were illustrated in Table (1). Plants were thinned twice the first after 10 days and the second after 21 days from sowing. Calcium super phosphate (15.5% P₂O₅) and potassium sulfate (48.5% K₂O) in the rate of 3.0 and 1.5 g/pot were added by broadcasting before sowing. Ammonium sulfate was added in two equal portions in the rate of 1.5 g/pot twice at 21 days from sowing and two weeks after the first one. Irrigation interval treatments were applied at 21 days till two weeks before harvesting.

Osmotic pressure determination

Osmotic pressure in fresh samples was determined according to the method described by Gusev (1960).

Chemical determinations

Two plants from each pot were collected, cleaned, dried at 70 °C in an electric oven for three days and ground in a stainless steel mill. The following determinations were investigated.

Nitrogen and protein determination

Digestion and N determination was done according to the methods described by micro Kejeldahel methods as described by **Pregel (1945)**.

Carbohydrate percentage determination

Total carbohydrate was determined according to the method of Smith, et al. (1956).

Oil percentage determination

Oil percentage was determined according to the methods described in the A.O.A.C. (1970).

Proline determination

Proline determination was done using the method described by: <u>Wren</u> and <u>Wiggall</u> (1965).

Collected Data was subjected to the proper statistical analyses as described by Snedecor and Cochran (1990).

RESULTS AND DISCUSSION

Table (1): Physical and chemical properties of soil under experimentation.

Physical properties							
C	'lay %	18.00					
Sa	and %	57.25					
5	Silt %	29.75					
Soil	Texture	San	dy loam				
Chemical analysis							
PH	H (1:3)	7.25					
EC	2. (1.53)	1.1 dS/m-1					
Available 1	macronutrients	Available micronutrients					
(ppm)						
Ν	189.10	В	3.42				
Р	3.14	Fe	15.14				
K	259.75	Mn	21.81				
Ca	65.15	Zn	1.18				
Mg	73.18	Cu	1.31				
		Al	0.78				

Growth and yield

Irrigation intervals: The plant height of irrigated plants every two weeks (67.8cm) exceeded those irrigated every one week or irrigated every three weeks. Number of green leaves/plant followed the same trend of plant height (Table, 2). Area of leaves was not affected by extension of irrigation interval, except for extension to three weeks. Total dry weight positively responded to GHS treatments when canola plants received water every 7 days. Prolonging irrigation to 21 days decreased dry mass with the first concentration (100 ppm glutathione) and showed slight increase with 200 ppm glutathione exceeding the control (Table 2).

Illustrated data in Table (3) showed the effect of irrigation interval on canola yield and its attributes, where the highest values were obtained when irrigating plants every two weeks. Seeds weight per plant increased by 36.0% and 387.10%, and number of seeds per pod increased by 4.95% and 82.03%, when irrigating plants every two weeks compared with one week and three weeks intervals, respectively. The drought stress occurs in the plant when the received water is less than water loss. This may results from either water over loss or reduction in the absorption or both cases (Kuchaki and Alizade, 1995). The drought stress affects different aspects of plant growth and causes reduction and delay of germination, reduction of shoots growth, and reduction of dry matter production. The reduction of osmotic potential and water potential, turgescence removing, and stoma closure as well as growth failure are symptoms of water stress (Singh and Patel, 1998). Gul and Ahmed (2004) noticed that vegetative growth recorded in term of plant height, number of leaves , number of branches and fresh biomass/plant was much reduced under 6 days intervals, it caused considerable reduction. Growth of plants depends on cell expansion and enlargement which is probably most sensitive physiological aspect of a plant to water deficit leading to reduce plant productivity (Larson, 1992).

	CIIC	Plant height, cm	leaves No.	Leaf	Fresh weight (g):			Dry weight (g):			
Irrigation intervals	ppm			area, mm ²	Ste	Leave	Total	Ste	Leav	Total	
					m	s		m	es	Total	
7	D.W.	47.3	9.67	31.50	10.1 2	28.02	38.14	0.70	1.81	2.51	
days	100	58.3	11.67	25.94	7.21	31.78	38.99	1.00	1.87	2.87	
	200	53.3	11.00	42.80	7.46	12.60	20.08	1.30	3.32	4.62	
14	D.W.	63.7	10.17	30.85	6.19	22.13	28.32	1.33	2.36	3.69	
14 dova	100	78.7	15.67	41.00	6.71	48.00	54.71	2.28	3.08	5.36	
uays	200	61.0	14.33	28.33	8.02	9.79	17.81	0.56	1.30	1.86	
21	D.W.	42.7	9.33	42.80	7.63	20.48	27.81	1.40	1.52	2.92	
21	100	51.3	8.67	28.33	6.06	37.11	43.17	0.39	1.31	1.70	
uays	200	45.0	11.33	16.78	5.15	19.47	24.62	0.75	1.52	2.27	
Mean values under	7	46.3	10.78	33.41	8.26	24.13	32.39	1.00	2.33	3.33	
the effect of	14	67.8	13.39	33.39	6.97	26.64	33.61	1.39	2.25	3.64	
Irrigation intervals	21	46.2	9.78	29.30	6.28	25.69	31.97	0.85	1.45	2.30	
Mean values under	D.W.	51.2	9.71	35.05	9.51	23.54	33.05	1.14	1.90	3.04	
the effect of glutathione	100	62.8	12.00	31.76	10.5 9	38.96	49.55	1.22	2.09	3.31	
rates	200	53.1	12.22	29.30	9.77	17.13	26.90	0.87	2.05	2.92	
LSD at 5% level	Ir.In.	9.72	2.58	N.S	N.S	N.S	8.69	N.S	N.S	1.37	
	GHS ·	7.05	1.72	N.S	N.S	1.96	8.01	N.S	1.19	N.S	
	Ir.In x GHS	N.S	2.98	14.76	N.S	3.40	13.86	N.S	N.S	3.34	

Table (2): Effect of glutathione and irrigation intervals on growth of canola.

Tuste (s). Encod of graduatione and filigation intervals on yield of canona									
Irrigation intervals	GHS ppm	No of pods/plant	Seeds weight/plant, g	No of seeds/pod	Weight of 100 seed, g				
7	D.W.	43.3	2.19	23.3	0.28				
/ deva	100	56.7	4.37	21.7	0.38				
uays	200	50.0	3.44	21.7	0.29				
14	D.W.	52.7	3.77	23.2	0.31				
14 deva	100	74.0	6.66	23.3	0.37				
uays	200	85.0	3.15	23.0	0.22				
21	D.W.	53.3	0.73	11.7	0.15				
	100	56.7	0.57	16.7	0.17				
uays	200	38.3	0.44	10.0	0.18				
Maan walnaa uu dan tha affaat	7	49.8	3.33	22.2	0.32				
of Immigation intervals	14	70.6	4.53	23.3	0.27				
of fifigation litter vals	21	49.4	0.93	12.8	0.17				
Maan walnaa uu dan tha affaat	D.W.	49.8	2.23	19.9	0.25				
Mean values under the effect	100	62.2	4.20	20.6	0.31				
or glutatinone rates	200	57.8	2.34	8.3	0.23				
	Ir.In.	N.S	0.81	3.90	0.013				
LSD at 5% level	GHS.	2.14	0.63	4.68	0.04				
	Ir.In x GHS	8.96	1.08	N.S	N.S				

Table (3): Effect of glutathione and irrigation intervals on yield of canola.

D.W. = Distilled water GHS.=Glutathione II.=Irrigation intervals

Growth and photosynthesis are two of the most important processes suppressed partially or completely by water stress (Kramer and Boyer, 1995). Several investigations concluded that this may be attributed to the effect on enzymes and oxidative defense (Niu, et al. 2013). Moisture stress affecting photosynthesis and carbohydrate building (Pinheiro and Chaves, 2011 and Tarighaleslami, et al. 2012).

Glutathione

Data in Table (2 & 3) for growth and yield showed that the enhancing effect of 100 ppm glutathione exceeded those induced by application of 200 ppm concentration, except for leaves number and area. Reduced glutathione, and reduced ascorbate, the two major water soluble antioxidants in photosynthetic and non-photosynthetic tissues, reacting directly or indirectly with reactive oxygen species, contribute to maintain the integrity of cell structures and the proper functions of various metabolic pathways (Konkline, et al. 1997). Eid, et al. (2011) noticed that glutathione increased plant height, number of branches, fresh and dry weight of herb and flowers and number of flowers in marigold (Tagetes erecta L.) plants.

Glutathione x irrigation intervals

Plant height and number of green leaves increased by GHS application (100ppm) under tested irrigation intervals. Area of leaves was reduced by application of 100 or 200 ppm GHS (Table 2). Total dry weight positively responded to GHS treatments when canola plants received water every 7 days and negatively responded to irrigation every 14 days but prolongation to 21 days dry mass decreased with 100ppm GHS and tended to increase with 200 ppm compared to the control. Moreover the highest total dry weight was attained by spraying plants with 100 ppm glutathione and irrigation every 14 days.

The interactive effects of irrigation intervals and glutathione spray amount on yield of canola plants. Data presented in (Table 3) showed that plant response to 100 ppm glutathione exceeded that of 200 ppm under 7 and 14 days irrigation intervals. However, under 21 days intervals, 100 ppm glutathione slightly affected number of pods but 200 ppm antioxidant concentration negatively affected this character (Table 3). Among the non-enzymatic compounds, glutathione and ascorbate are essential plant metabolites that regulate major cell functions and play a pivotal role in antioxidant defense (Noctor and Foyer, 1998). Exposure of plants to unfavorable conditions such as drought, high temperature or salinity can increase the production of reactive oxygen species (ROS) single oxygen, hydrogen peroxide, superoxide radicals, and hydroxyl radicals to protect themselves against these toxic oxygen intermediates. Plants employ defense that included the enzymes such as suproxide dismutase, catalases, ascorbate peroxidases, glutathione-s-transferases and glutathione reductase that catalyze the scavenging of ROS (Rexas, et al. 2000).

Carbohydrate, oil and protein

Irrigation intervals

Protein and carbohydrate percentages were reduced by extending the irrigation interval, meanwhile, oil percentage increased with 14 days intervals and decreased with 21 days intervals to be less than the control (Table 4). This means that extension of irrigation interval adverse markedly the carbohydrate and protein building. Such result was confirmed by Pinheiro and Chaves (2011) and Tarighaleslami, et al. (2012). The plant reacts to water deficit with a rapid closure of stomata to avoid further loss of water through transpiration. As a consequence, the diffusion of CO₂ into the leaf is restricted, less water in tissues and this in turn affected the carbohydrates building (Kramer and Boyer, 1995).

Glutathione: It was clear that carbohydrate content increased as sprayed with GHS but protein and oil percentages were not affected (Table 4). Eid, et al. (2011) found that application of glutathione (100 or 200 ppm) was effective in increasing carbohydrate percentage and amino acid concentrations.

Glutathione x irrigation intervals

Spraying plants with 100 ppm GHS under 7 days irrigation intervals showed the highest value of carbohydrate .Oil percentage only increased with application of 100 and 200 ppm GHS under 7 days intervals.

Irrigation intervals	GHS ppm	Carboh -ydrate	Protein %	Oil %	Proline ppm	Osmoti c pressur e
7		52.7	22.85	23	2.48	5.12
/ deve	D.W.	86.9	24.81	25	4.69	6.60
uays	100	64.9	24.69	27	2.40	5.67
14	200	56.0	23.38	31	1.94	5.36
14 deva	D.W.	59.8	24.31	31	3.01	4.77
uays	100	55.5	23.33	27	7.72	6.61
21	200	59.7	20.87	22	1.09	6.07
21	D.W.	60.0	23.88	21	2.64	6.20
uays	100	65.0	23.44	21	3.21	5.30
Moon values under the effect of Invigation	200	68.2	24.12	25.0	3.19	6.46
intervale	7	57.1	23.69	29.7	4.22	5.58
Intervais	14	61.6	22.78	21.3	2.31	5.86
Moon values under the effect of slutethions	21	56.1	23.45	25.3	1.84	5.52
wreat values under the effect of glutathione	D.W.	68.9	24.33	25.7	3.45	5.86
Tates	100	61.8	23.84	25.0	4.44	6.53

Table (4):	Effect of	glutathione and	l irrigation	intervals on	chemical	constituents of	canola	plants
		O						

D.W. =Distilled water GHS.=Glutathione II.=Irrigation intervals

Osmotic pressure and proline

Irrigation intervals

Data in Table (4) showed the decrease in osmotic pressure with the increase in periods between irrigations. Huang and Redman, 1995) mentioned that osmotic adjustment play a crucial role in plant adaptation to drought. Glutathione

Application of glutathione showed slight increase in osmotic pressure of canola plants and 100 ppm GHS increased proline under 14 days irrigation interval Table (4). Water uptake is enhanced by the accumulation of solutes to lower the tissue water potential and by improving root growth, and water loss through evaporation is limited by closing stomata, restricting shoot growth and accelerating leaf senescence (Nakashima, et al. 2009). Exogenous amino acids application caused significant increase in proline contents (Burbulis, et al. 2013).

Glutathione x irrigation intervals

The response of OP to application of GHS was similar under 4 and 21 days between irrigations but under 7 days intervals was different which decreased with the 100 ppm treatment (Table 4).. Glutathione is one of the major endogenous antioxidants in plants known to play an important role in plant defense mechanisms. Glutathione Stransferase is a GSH dependent detoxifying enzyme in plants, which catalyzes the conjugation of GSH (Yilmaz, 2006). The response of OP to application of GhS was similar under 4 and 21 days between irrigations but under 7

days intervals was different which decreased with the 100 ppm treatment and tended to increase to be more than that was shown with control plants. The foliar spray of GB could markedly alleviate the water stress (Cao, et al. 2013).

REFERENCES

A.O.A.C. (1970). Association Agricultural Washington D C.200 Official Methods of Analysis, 12th ed., 44:94-117. Bray, E.A. (1997). Plant response to water deficit. Trends Plant Sci., 2:8-54.

Burbulis, N.; Jonytienė, V. and Blinstrubienė, A. (2013). Effect of exogenous amino acids on electrolyte leakage in rapeseed shoots cultured in vitro. Food, Agriculture and Environment (JFAE), 11 Issue 3&4: 2119-2122.

<u>Bybordi</u>, A. and <u>Ebrahimian</u>, E. (2013). Growth, yield and quality components of canola fertilized with urea and Zeolite. Communications in Soil Science and Plant Analysis, 44, Issue 19: 2896-2915.

Campos, P. S. (1998). Effects of water stress on photosynthetic performance and membrane integrity in Vigna spp. The role of membrane lipids in drought tolerance. Ph.D. dissertation. Universidade Nova de Lisboa, Lisboa.

Cao, F.; Liu, L.; Ibrahim, W.; Cai, Y. and Wu, F. (2013) Alleviating Effects of Exogenous Glutathione, Glycinebetaine, Brassinosteroids and Salicylic Acid on Cadmium Toxicity in Rice Seedlings (Oryza Sativa). Agrotechnol, 2: 107.

<u>Chen</u>, J.; <u>Jiang</u>, H.; <u>Hsieh</u>, E.; <u>Chen</u>, H.; <u>Chien</u>, C.; <u>Hsieh</u>, H., and <u>Lin</u>, T.(2012). Drought and Salt Stress Tolerance of an Arabidopsis Glutathione S-Transferase U17 Knockout Mutant Are Attributed to the Combined Effect of Glutathione and Abscisic Acid. Plant Physiol., 158(1): 340–351.

Coleman, J.O.D.; Blake-Kalff, M.M.A. and Davies, T.G.E. (1997). Detoxification of xenobiotics by plants: chemical modification and vacuolar compartmentation. Trends Plant Sci., 2: 144–151.

Eid, R. M.; Lobna M.; Taha, M. and Ibrahim, S.M. (2011). Alleviation of adverse effect of salinity in growth, and chemical constituents of margold plants. J. Appl. Sci. Res., 7(5): 714-72

Gallab, K.H. and Sharaan, A.N.(2002).Selection in canola (Brassica nupas L) germplasm under newly reclaimed lands; salt tolerant selections.. Egypt. J. Plant Breeding, 6(2):15-30.

Glantz, M.H. (1994). Drought, Desertification and food production. In drought follows the Plow. Eds, M.H. Glantz pp.7-32. Cambirdge Univ, pr.

Gusev, N.A. (1960). Some methods for study plant water relations. Akad. Nauke. S.S.S.R. Lenengrad.

Huang , J. and Redman, R.E. (1995). Physiological response of canola and wild mustard to salinity and contrasting calcium supply . J. of Plant Nutr., 18(9): 1931-1949.

James, V. A. and Davis, D.G.(2004). Abiotic stress alters transcript profiles and activity of glutathione S-transferase, glutathione peroxidase, and glutathione reductase in Ephorpia esula. Physiologia Plantarum, 120:421-433.

Konkline, P. L.; Palanca, J.; Last.R .L .and Simeroff, N. (1997). L ascorbic acid metabolism in ascorbate deficient in Arapidopsis mutant vtc1.Plant Physiol., 115: 1277-1285.

Kramer, B.J. and Boyer, J.S. (1995). Water Relations of plant and soil. Acad. Press INC. San Diego.

Kuchaki, A. and Alizade, A.(1995). Agriculture principles in dry regions. Razavi Press, 260.

Lareson, K.L. (1992). Drought injury and resistance of crop plants. In Physiological Aspects of Dry Land Farmin. (Eds) S.U. Gupta pp. 147-162. Oxford & IBH publishing Com. Pvt Ltd., Newdelhi, India.

Mekki, B. (2013) Yield and quality traits of some canola varieties grown in newly reclaimed soils. World appl.Sci.J., 25(2):258-263.

Nakashima, K.; Ito, Y.; Yamaguchi-Shinozaki, K. (2009) Transcriptional regulatory networks in response to abiotic stresses in arabidopsis and grasses. Plant Physiol, 149: 88-95.

Niu, Y.; Wang Y. and Li, P. (2013). Drought stress induce oxidative stress and antioxidant defense and ascorbate deficient vtc1 mutant of Arabidopsis thalianaa. Acta Physiol.Plant., 35: 1189-1200.

Noctor, G. and Foyer, C.H. (1998) Ascorbate and glutathione: keeping active oxygen under control. Plant Biol., 49: 249-279.

Pinheiro, C. and Chaves, M.M. (2011). Photosynthesis and drought: can we make metabolic connections from available data?. J. Exp. Bot., 62 (3): 869-882.

Pompella, A.; Viavikis, A.and Paolicche, A.(2003). The change faces of glutathione a cellular protagonist. Biochem. Pharma., 66:1499-1503.

Pregel, F., 1945. Quantitive organic micro analysis. 4 Ed. J. A. Churchill Ltd., London, pp: 53.

Rexas, V.P.; Lodhi, S.A.; Garrett, D.A.; Mahan, J.R.and Allen, R.G. (2000). Stress tolerance in transgenic tobacco seedlings that over express glutathione-s-transferase/glutathione peroxidase. Plant cell Physiology, 41 Issue 11:1229-1234. Ramanjulu, D. B. (2002).Drought- and desiccation-induced modulation of gene expression in plants. Plant Cell Environ., 25: 141–415.

Sakr, M.T. and <u>Arafa</u>, A.A. (2009). Effect of some antioxidants on canola plants grown under soil salt stress condition. <u>Pakistan Journal of Biological Sciences</u>, 12(7):582-588.

Shabani, A.; Sapaskhah, A.R. and Kamgar-Haghighi, A.A. (2013). Responses of production compounds of rape seed as influenced by deficit irrigation, water salinity and planting method. Intr. J. of Plant Prod., 7(2): 313-340.

Singh, J. and Patel, A.L. (1998). Water statues, gaseous exchange, prolin accumulation and yield of wheat in response to water stress. Annual of Biology Ludhiana, 12: 77-81.

Smith, F.M.; Gilles, A.; Amilton, D.K. and Geders, P.a. (1956). Colormetric methods for determination of sugar and related substances. Ann. Chem., 28:350-356.

Snedecor, G.W., Cochran, W.G. 1990. Statistical Methods. 11th Ed. Iowa State Univ., Press. Ames, Iowa, USA.

Tarighaleslami, M.; Zarghami, R. Mashhadi, M.; Boojar, K. and Oveysi, M. (2012). Effects of Drought Stress and Different Nitrogen Levels on Morphological Traits of Proline in Leaf and Protein of Corn Seed (Zea mays L.). American-Eurasian J. Agric. & Environ. Sci., 12 (1): 49-56.

Ullah, F.; Bano, A. and Nosheen, A. (2012). Effect of plant growth regulators on growth and oil quality of canola under drought stress. Pak. J. of Plant Bot., 44(6): 1873-1880.

Wang, W.; Vinocur, B. and Altman, A. (2003). Plant responses to drought, salinity and extreme temperatures: towards genetic engineering forstress tolerance. Planta., 218:1-14.

Wiess, E.A. (1983).Oil seed crop . pp. 61-215. Longman Group Limited.

Wren, J.J. and <u>Wiggall</u>, P.H.(1965). An improved colorimetric method for the determination of proline in the presence of other ninhydrin-positive compounds. Biochem. J., 94: 216-220.

Yılmaz, C. (2006). Glutathione s-transferase activity and glutathione levels in drought stresses Pinus brutal Ten trees growing in Ankra. M.Sc. Thesis Graduate School of Natural and Applied Sciences. Department of Biochemistry, Ankra, Turky.